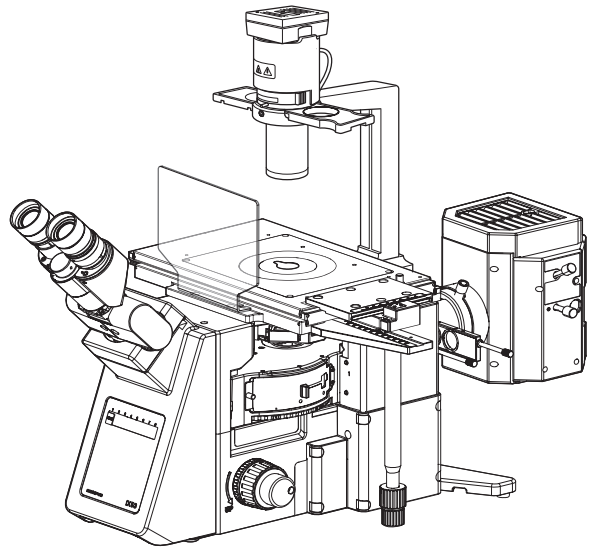


OLYMPUS®



INSTRUCTIONS

IX53

INVERTED MICROSCOPE

This instruction manual is for the Olympus inverted microscope model IX53.

To ensure the safety, obtain optimum performance and to familiarize yourself fully with the use of this microscope, we recommend that you study this manual thoroughly before operating the microscope.

Retain this instruction manual in an easily accessible place near the work desk for future reference.



AX8169



In accordance with European Directive 2002/96/EC on Waste Electrical and Electronic Equipment, this symbol indicates that the product must not be disposed of as unsorted municipal waste, but should be collected separately.

Refer to your local Olympus distributor in EU for return and/or collection systems available in your country.

IMPORTANT – Be sure to read this section for safe use of the equipment. – 1-5

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Caution

If the equipment is used in a manner not specified by this manual, the safety of the user may be imperiled. In addition, the equipment may also be damaged. Always use the equipment as outlined in this instruction manual.

The following symbols are used to set off text in this instruction manual.

CAUTION : Indicates a potentially hazardous situation which, if not avoided, may result in minor or moderate injury or damage to the equipment or other property. It may also be used to alert against unsafe practices.

© : Indicates commentary (for ease of operation and maintenance).

IMPORTANT

This microscope employs UIS2 optical design.

For the applicable modules, please consult Olympus or refer to the latest brochures or Olympus website. Less than optimum performance may result if inappropriate module combinations are used.

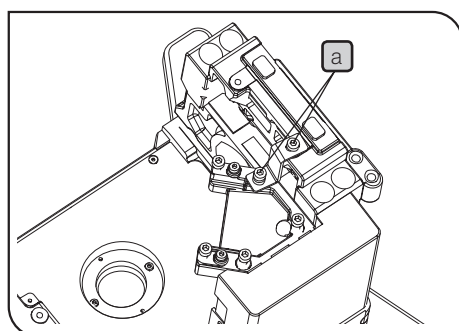
Configuration of Instruction Manuals

Since this microscope is expandable to a variety of systems, separate instruction manuals are prepared so that the user has to read only the manuals according to the user's own system.

Manual Name	Main contents
IX53 (this instruction manual)	Observation procedures including transmitted light brightfield, phase contrast
IX53 REFLECTED FLUORESCENCE SYSTEM	Reflected light fluorescence observation
IX2-GS	Gliding stage
TL4	Power supply for 30 W halogen lamp
TH4	Power supply for 100 W halogen lamp
IX2-MLWCD	Mid-Long working distance condenser

Releasing the Transport Lock

Be sure to work on the following tasks first after finishing the unpacking.



Releasing the transport lock of the focusing mechanism

CAUTION Never attempt to rotate the focusing knob without removing the clamping plate. Otherwise, the focusing mechanism may be damaged.

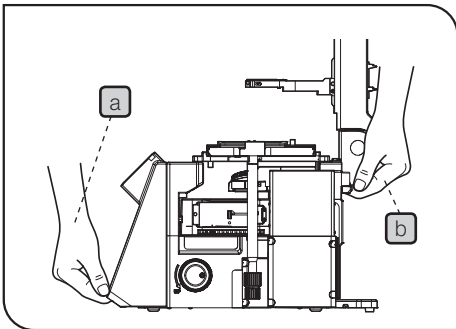
Loosen and remove the screws **a** of the clamping plate using the Allen wrench provided with the microscope.

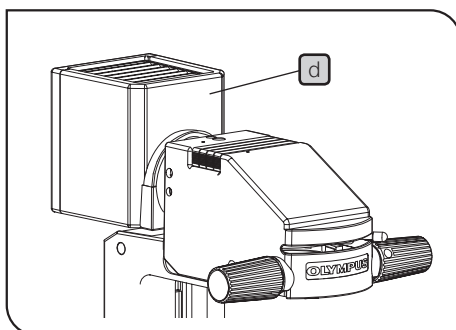
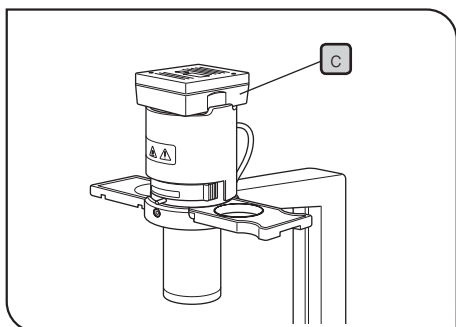
- © Keep the clamping plate and screws in a safe place because it will be used again when the microscope is transported next time.



SAFETY PRECAUTIONS

1. If potentially infectious samples may be observed, use protective gloves or other protective means to prevent the skin from contacting with samples directly.
After observation, be sure to clean the portion contacted with samples.
 - Moving this device is accompanied with the risk of dropping the samples. Be sure to remove the samples before moving this device.
 - In case the samples are damaged by erroneous operation, promptly take the infection prevention measures.
 - Follow the procedures described in "Handling Precautions" (see page 5) prior to using the accessories of this device. Otherwise, the stability of the device will be lost and the dropped samples will cause the possibility of infection.
 - When you maintain the device which may have contacted with potentially infectious samples, be sure to wear protectors such as gloves, or clean the device prior to operation.
 - Before disposing of the device contacted with potentially infectious samples, be sure to follow the regulations and rules of your local government.
2. Be careful not to have your hand caught between the bottom of the revolving nosepiece and the microscope frame.
3. The microscope is not provided with a waterproof mechanism. Therefore, if culture liquid or water is spilt on the stage, revolving nosepiece or microscope frame, damage to the equipment or an electrical shock may result. Immediately wipe the liquid or water off if it is spilt on them.
4. The microscope is not provided with a dust-proof mechanism.
5. Install the microscope on a sturdy, level table or bench.
6. When moving the microscope, remove the modules that may drop including the specimen from the microscope in advance, then carefully carry the microscope frame by holding the base (front edge) **a** and the grasping part below the illumination column **b**.
7. When carrying the microscope to a different place, it is also recommended to disconnect all cables and modules from the microscope frame.
When transporting it, also engage the transport lock mechanisms and package it sufficiently.
Also be careful against slipping of hands during carrying.
8. Damage to the microscope will occur if you grasp it by other parts including the stage, focusing knob, etc.
9. This microscope is not equipped with laser safety mechanisms. The user should assume liabilities for any consequence of user modification including introduction of the use of a laser beam.
10. If laser equipment is attached to the product you have purchased, this Instruction Manual is not effective. Follow the Instruction Manual provided with the laser equipment to be attached.
11. The surfaces of the lamp housing will become extremely hot during operation. When installing the microscope, make sure to allow ample free space (10 cm or more) around and in particular above the lamp housing.
12. When installing the microscope, route the power cord away from the lamp housing. Should the power cord come in contact with a hot part, the power cord could melt and cause electric shock.





13. After operation or in case of abnormality, be sure to disconnect the power cord from the connector on the products or from the outlet.
14. To avoid potential shock hazards and burns when replacing the lamp bulb, set the main switch to "O" (OFF) then disconnect the power cord from the wall outlet in advance. Whenever you replace the bulb during use or right after use, allow the lamp housing **c** or **d** and bulb to cool before touching.





Designated bulb	6V30WHAL (PHILIPS 5761)
(IX2-ILL30)	

Designated bulb	12V100WHAL-L (PHILIPS 7724)
(IX3-ILL)	

15. Do not permit tools or metal fragments to get into air vents, or other apertures. Doing so could cause failure of the microscope or electric shock to the user.
16. The standard service life of the lamp housing is eight (8) years of use or 20,000 hours of total power ON period, whichever is the shorter period. For details, see the Inspection Sheet on page 50-51.
17. During the fluorescent observation, be sure to attach the anti-glare board to the microscope to protect your eyes.
18. If you feel bright during observation through eyepieces, darken the illumination light by adjusting the brightness of the light source or using the ND filter. Use the equipment according to the regulations for workers health and safety, if any.
 - Europe: 2006/25/EC Directive for protecting workers from risks caused by physical factors (artificial optical radiation).
19. Do not look directly at the light from the objective lens or the specular reflection light of the specimen.
20. Be careful invisible wavelength light (ultraviolet or infrared) may be emitted depending on illumination methods.
21. If the skin is exposed to the light from the objective lens for a long time, it may cause burns. Be sure to avoid it.
22. Do not place flammable gas or liquid close to the light from the objective lens. It may cause fire.
23. This device complies with the emission and immunity requirements described in IEC61326 series.
24. The electromagnetic environment should be evaluated prior to operation of this device. Do not use this device in close proximity to the sources of strong electromagnetic radiation to prevent interference with the proper operation.
25. Always use the power cord provided by Olympus. If no power cord is provided, please select the proper power cord by referring to the section "PROPER SELECTION OF THE POWER CORD" at the end of this instruction manual. If the proper power cord is not used, the safety and EMC performance of the device can not be assured.
26. Always connect the power cord correctly and ensure that the grounding terminal of the device and that of the wall outlet are properly connected. If the device is not grounded, our intended electric safety and EMC performance of the device can not be assured.





Safety Symbols

The following symbols are found on the microscope. Study the meaning of the symbols and always use the equipment in the safest possible manner.

Symbol	Explanation
	Indicates that the surface becomes hot, and should not be touched with bare hands.
	Indicates a non-specific general hazard. Follow the description given after this symbol or in the instruction manual.
	Indicates that the main switch is ON.
	Indicates that the main switch is OFF.

Caution labels

Caution labels are placed at parts where special precaution is required when handling and using the microscope. Always pay attentions to the caution labels.

Positions of caution labels	Lamp housing	 
	Front face of the illumination column IX2-ILL30	 

If the caution label becomes dirty or is peeled off, contact Olympus for replacement.

1 Intended use

This device has been designed to be used to observe magnified images of specimens in various routine work and research applications.

Do not use this device for any purpose other than its intended use.



This product complies with the requirements of directive 98/79/EC concerning in vitro diagnostic medical devices. CE marking means the conformity to the directive.

USA: CAUTION:

Federal law restricts this device to sale by or on the order of an appropriately licensed healthcare practitioner.

2 Handling Precautions

1. These products are precision instruments. Handle them with care and avoid subjecting them to sudden or severe impact and also connect the cables gently.
2. Do not use the microscope where it is subjected to direct sunlight, high temperature and humidity, dust or vibrations. (For operating conditions, see Chapter 8, "SPECIFICATIONS" on page 40.)
3. When attaching or detaching any accessory, make sure to proceed in a condition that nothing is attached on the left side port.
4. Do not disassemble any part of the microscope. Doing so could cause failure of the microscope.
5. Before disposing of this product, be sure to follow the regulations and rules of your local government.

3 Maintenance and Storage

1. Do not leave stains or fingerprints on the lenses and filters. Blow away dust with a commercially available blower and gently wipe the lens or filter with a piece of cleaning paper (or clean gauze).
For wiping fingerprints and oil stains, use a piece of cleaning paper moistened with commercially available absolute alcohol.

CAUTION

Since the absolute alcohol is highly flammable, it must be handled carefully. Be sure to keep it away from open flames or potential sources of electrical sparks --- for example, electrical equipment that is being switched on or off, which could cause ignition of a fire.

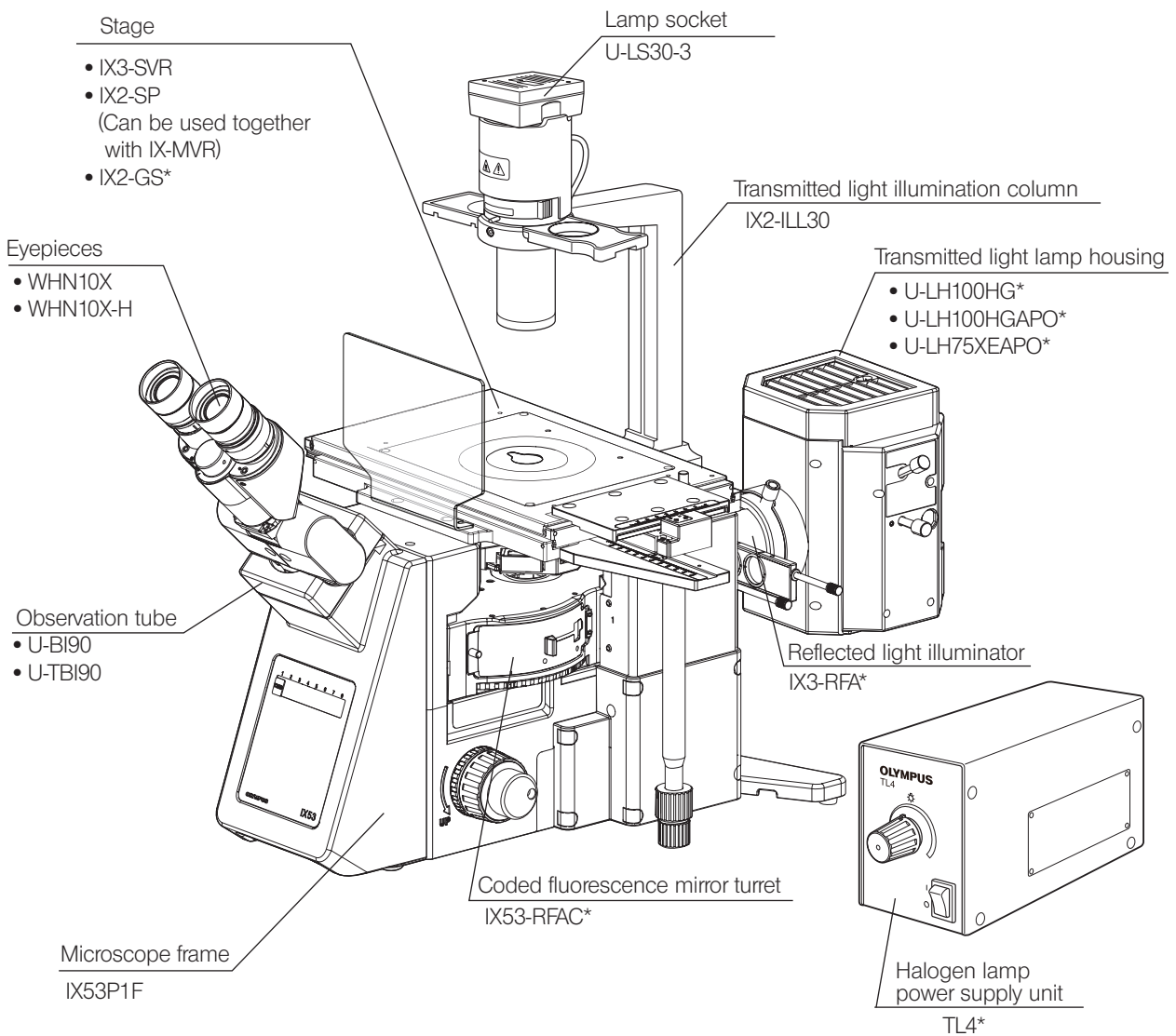
Also remember to always use absolute alcohol only in a well-ventilated room.

2. Do not use organic solvents, which cause painted and plastic parts to deteriorate. Do not use organic solvents to clean device components other than the glass components. To clean them, use a lint-free, soft cloth slightly moistened with a diluted neutral detergent.
3. This microscope is not provided with a dust-proof mechanism. When not using the microscope, make sure to set the main switch to "O" (OFF), confirm that the lamp housing is cool enough and cover the microscope with the provided dust cover.

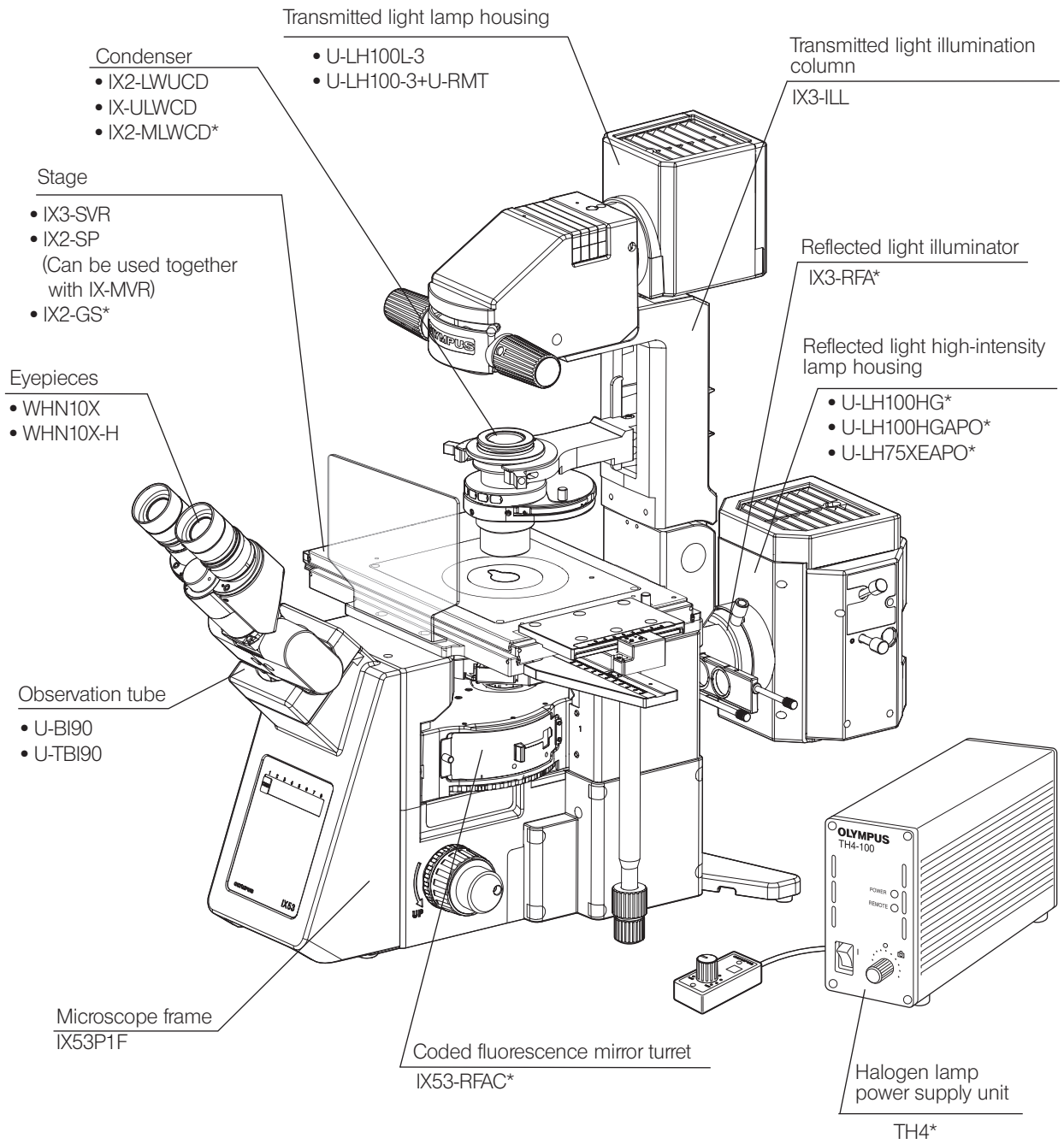
1 MODULE NOMENCLATURE

◎ The modules shown below are only the basic modules. As there are other modules which can be combined with the microscope but are not shown below, please also refer to the latest Olympus brochures or your dealer.
For information on the modules marked with “*”, refer to their instruction manuals.

with the IX2-ILL30

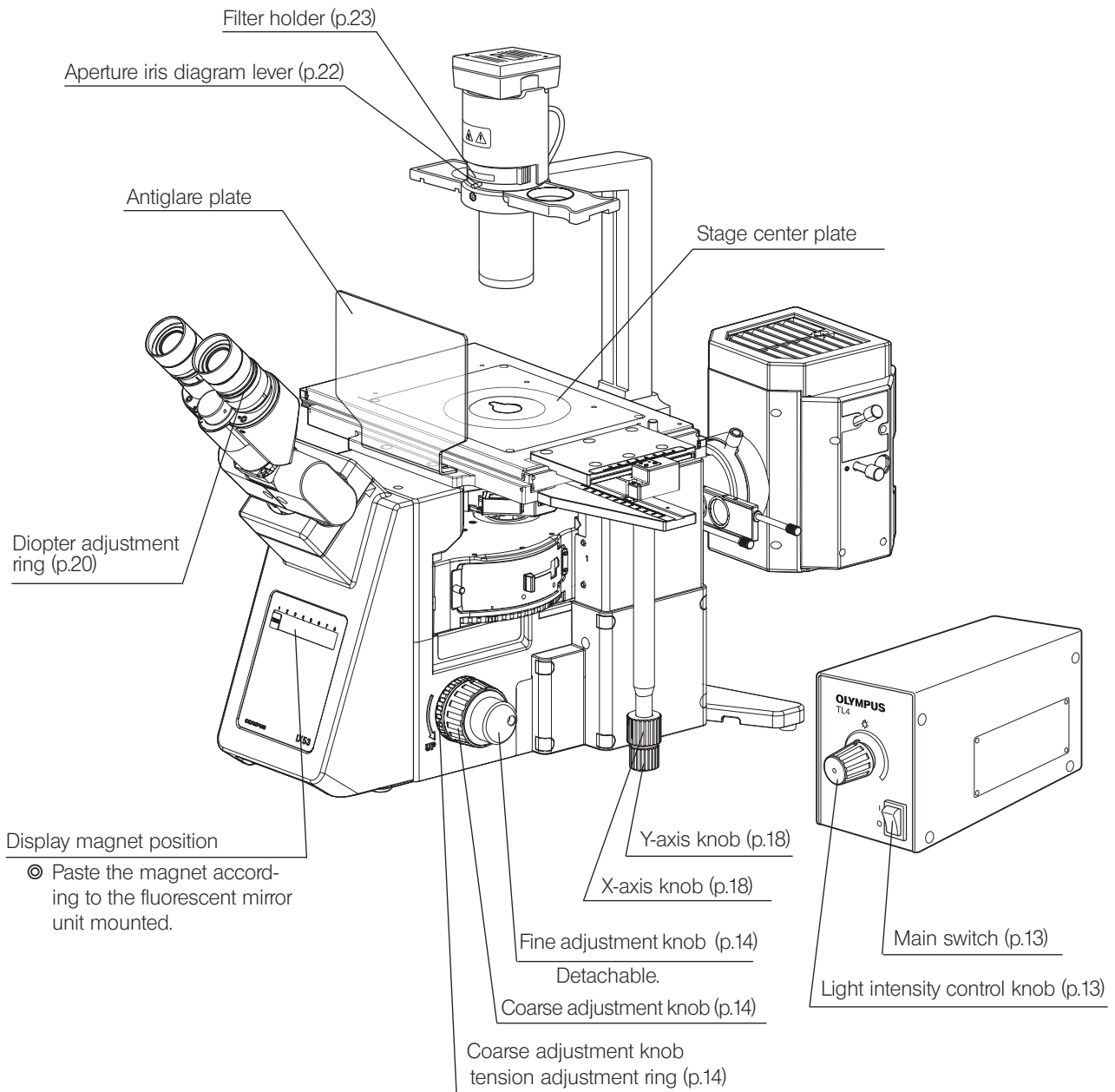


with the IX3-ILL

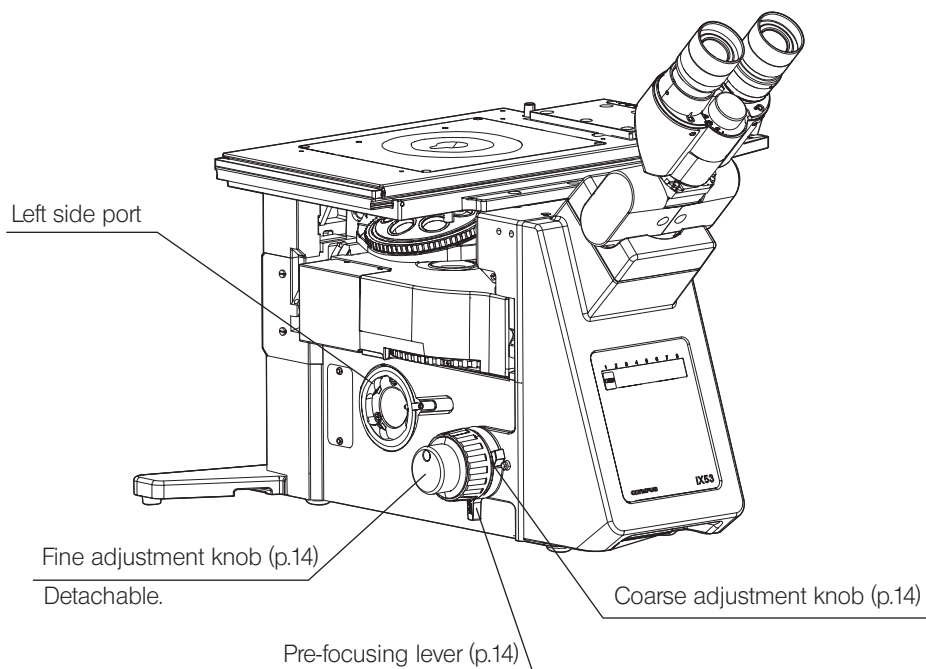


2 NOMENCLATURE

☉ If you have not yet assembled the microscope, read Chapter 9, "ASSEMBLY" (pages 41 to 49).



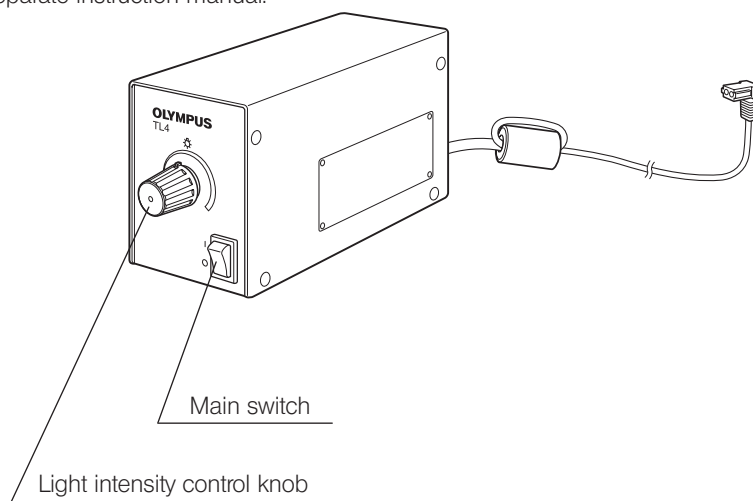
Left Side View of Microscope Frame



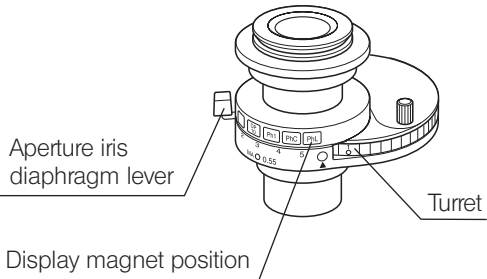
Other Modules

Halogen lamp Power Supply Unit TL4

- © The applicable halogen bulb is the 6V30WHAL
For details, refer to the separate instruction manual.

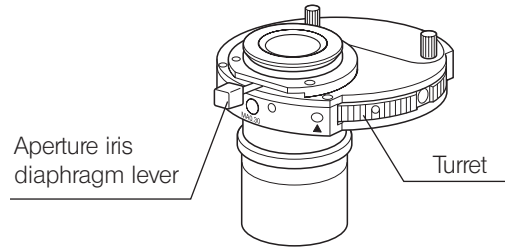


**LWD Universal Condenser
IX2-LWUCD**

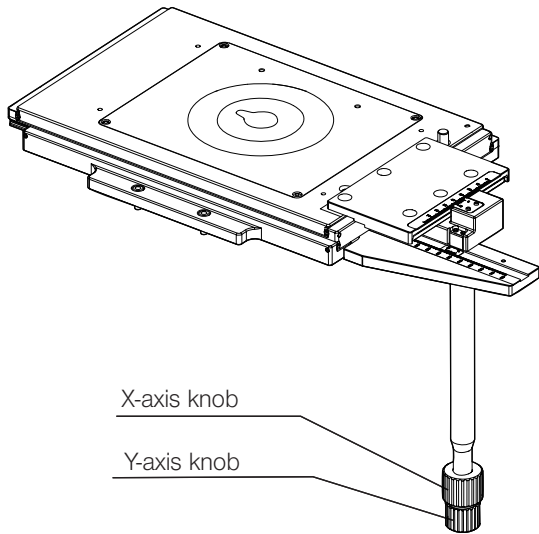


- ◎ Paste the magnet according to the optical element attached to the condenser.

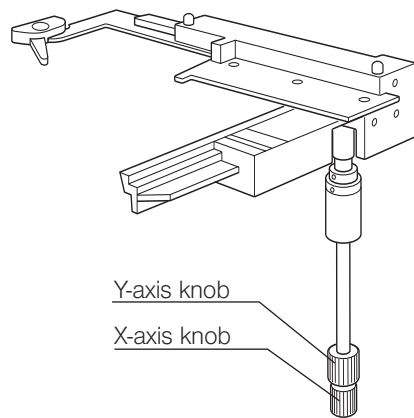
**ULWCD Condenser
IX-ULWCD**



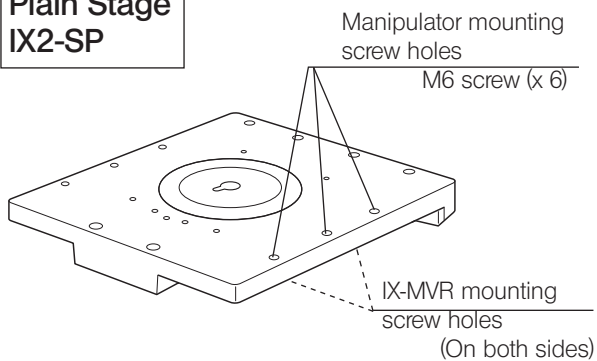
**Mechanical stage with right handle
IX3-SVR**



**Mechanical Stage
IX-MVR**



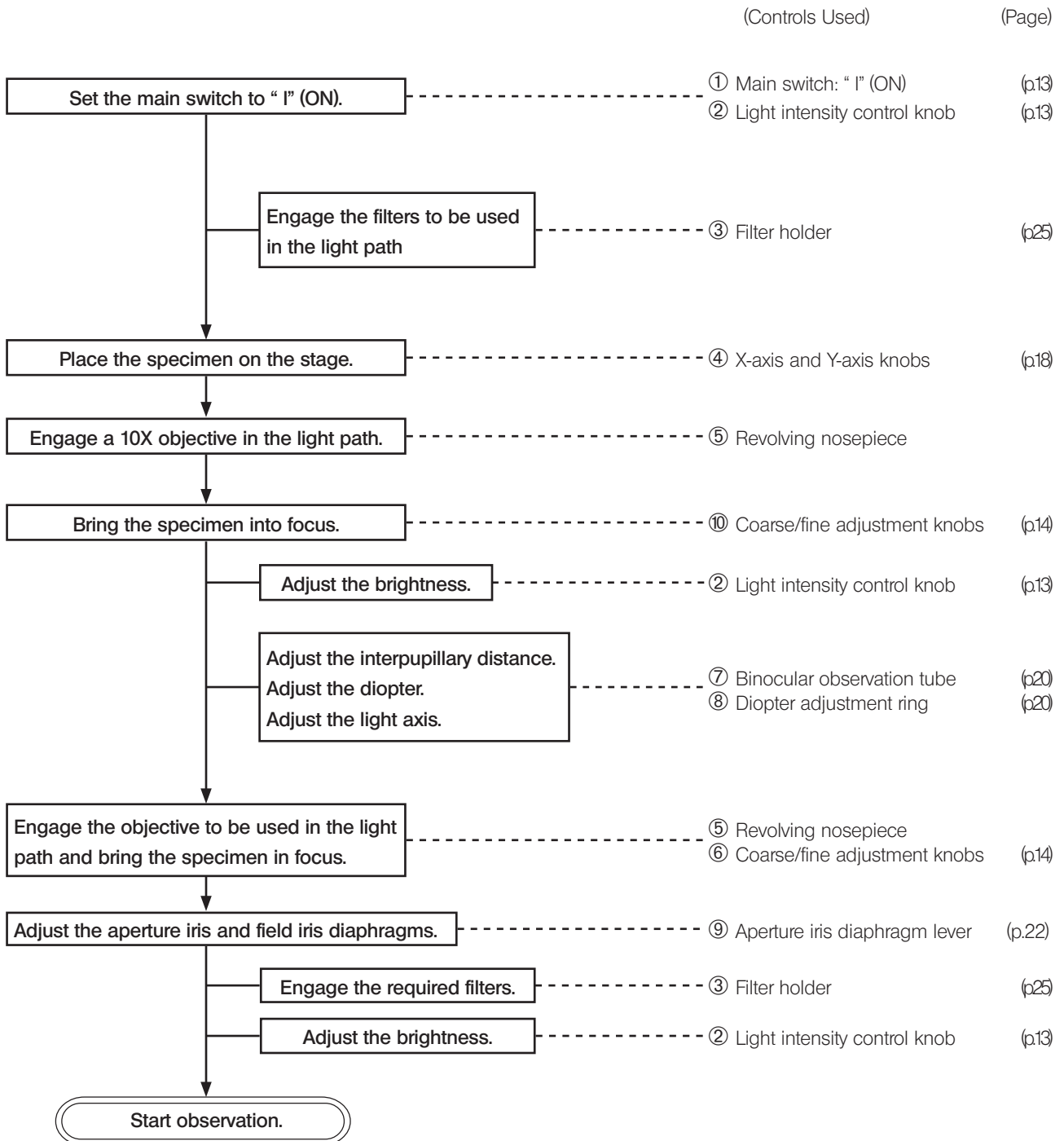
**Plain Stage
IX2-SP**

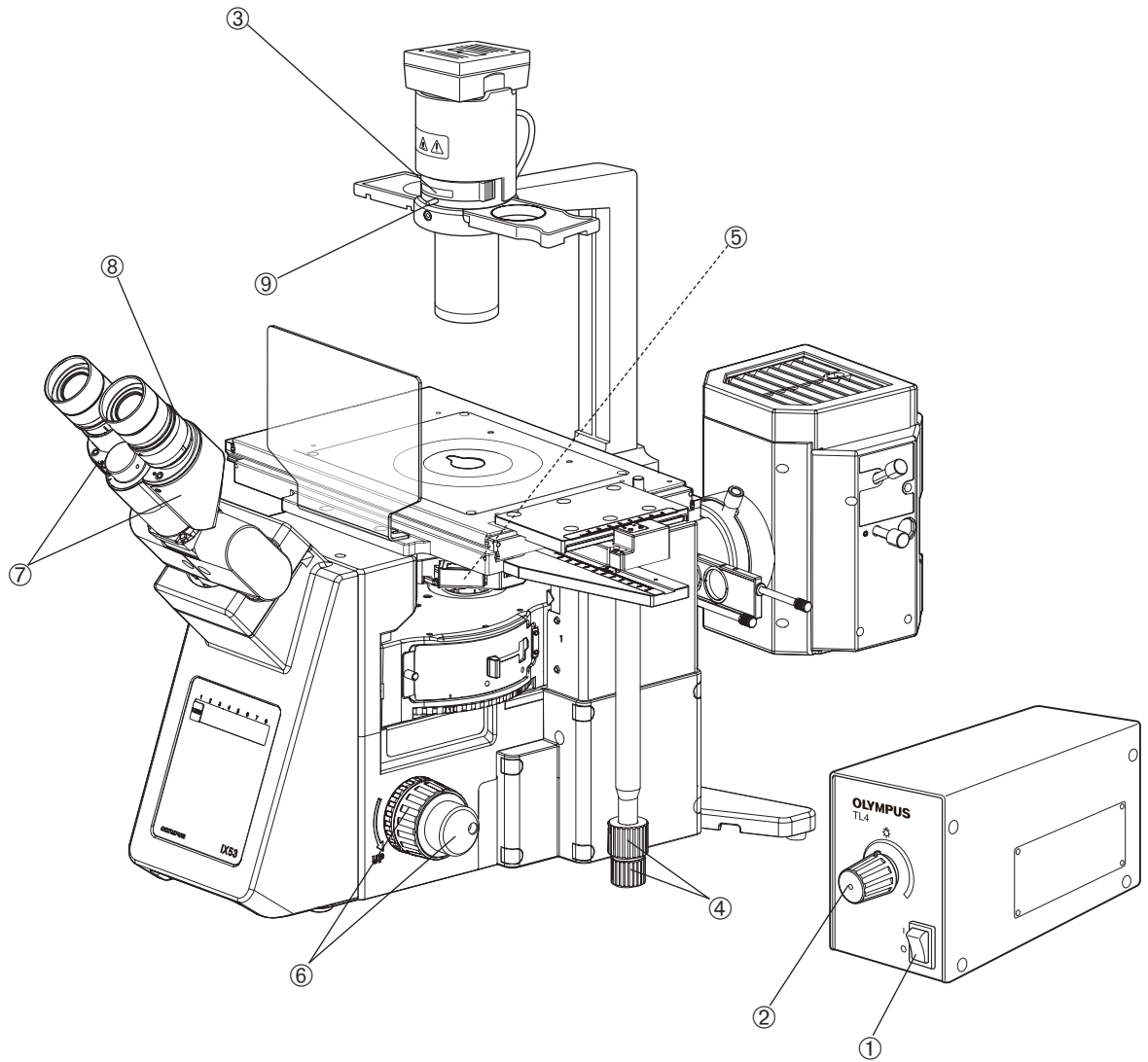


3 TRANSMITTED LIGHT BRIGHTFIELD OBSERVATION PROCEDURE

© The following flow shows the operating procedure for the transmitted light brightfield observation which is the basic observation method of this microscope. The operating procedures for phase contrast observation will be described separately in Chapter 5, "OTHER OBSERVATION METHODS" on page 31.

For the fluorescence observation, refer to the separate instruction manual entitled "Manual / Motorized Reflected Fluorescence System".





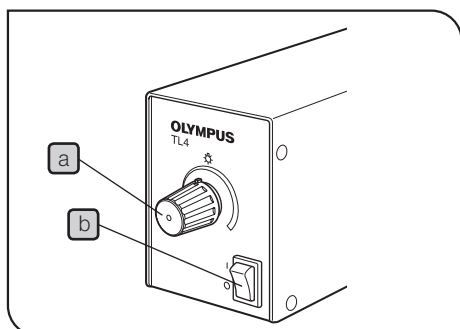
© Make a photocopy of the observation procedure pages and post it near your microscope.

4 USING THE CONTROLS

4-1 Power Supply Unit

1 Turning Power On, Adjusting the Brightness


TL4/TH4

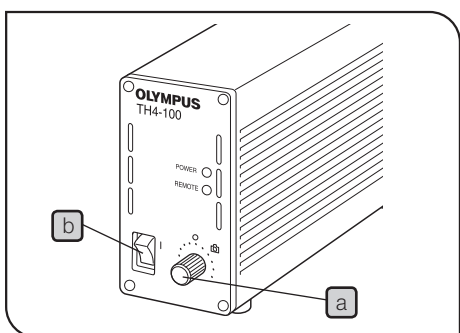


TL4

1 Make sure that the light intensity control knob **a** is in the MIN (minimum intensity) position and set the main switch **b** to "ON".

2 Rotate the knob **a** toward MAX (maximum intensity) to increase the intensity and the illumination brightness.

⊙ With the TH4, the position (approx. 9 V) marked  indicates the position where the daylight illumination suitable for photomicrography is obtained when the 45LBD filter is engaged in the light path.



TH4

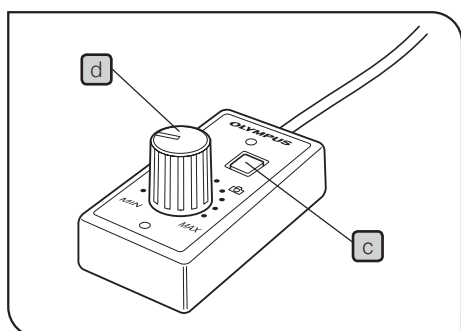
When the TH4-HS Hand Switch is Used

⊙ The illumination brightness can be adjusted from the hand switch.

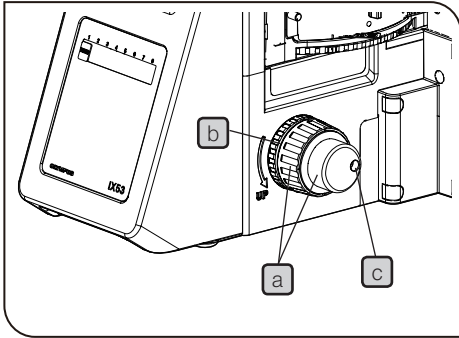
The hand switch is provided with double-sided adhesive tape, so it can be adhered to an easy-to-use position.

1 Press the transmitted light ON-OFF button **c** (so that the button is illuminated) and adjust the brightness with the light intensity control knob **d**.

2 To turn the lamp OFF, set the transmitted light ON-OFF button **c** to OFF.



4-2 Focusing Block



1 Rotation Direction of the Coarse/Fine Adjustment Knobs

- ⊙ Rotating the coarse or fine focus adjustment knob **a** toward the front (in the direction of the arrow) raises the objective and toward the rear (opposite direction) lowers the objective.

2 Adjusting the Coarse Adjustment Knob Tension

- ⊙ Always use the rotation tension adjustment ring **b** to control the rotation tension of the coarse adjustment knob.

The tension of the coarse adjustment knob has been pre-adjusted to optimum tension, but this can be changed as required. Turn the rotation tension adjustment ring **b** in the direction of the arrow to decrease the knob's tension and in the opposite direction to increase it.

3 Detaching the Fine Adjustment Knob

- ⊙ The fine adjustment knob is designed detachable in order to prevent interference between the knob and the operator's hand manipulating the X- and Y-axis knobs.
- Loosen the clamping screw **c** using the Allen screwdriver and remove the fine adjustment knob.
- After detaching, the fine adjustment knob is hollowed to facilitate manipulation with a fingertip.

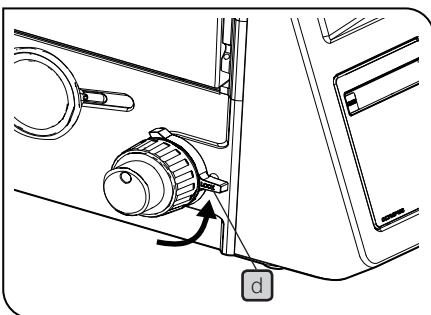
4 Pre-focusing Lever

- ⊙ The pre-focusing lever prevents collision between the specimen and objective and simplifies the focusing operation.

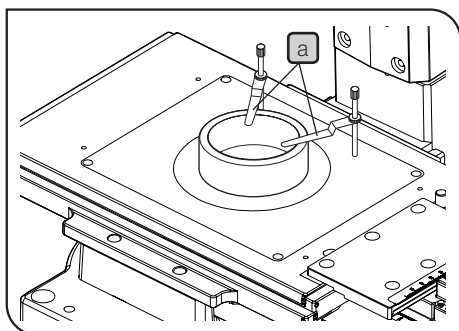
After bringing the specimen into approximate focus with the coarse adjustment knob, turn the pre-focusing lever **d** in the direction of the arrow to lock it. Hereafter, the upper limit of the coarse adjustment will be limited at the position where the lever is locked.

When bringing a specimen in focus, approximate focus can be obtained by simply raising the coarse adjustment to the stop position so all you have to do additionally is control the fine adjustment knob.

- ⊙ The focusing function using the fine adjustment knob is not limited.



4-3 Stage



1 Placing the Specimen

Place the specimen on the center of the stage.

- ⊙ If the specimen is prone to slide on the stage, attach the stage clips (IX-SCL) **a** and clamp the specimen down with the clips.

With the mechanical stage with right handle IX3-SVR

For IX3-SVR, in addition to the holder for the round stage center plate, following sample holders corresponding to each sample can be attached.

- IX3-HOW : Microplate holder
- IX3-HOS : Slide holder
- IX3-HO35D : Dish holder

See page 45 for assembly.

CAUTION The sample holder fixes the specimen to reproduce the specimen position. Do not push up the specimen by the objective lens. The specimen may be popped out.

IX3-HOW

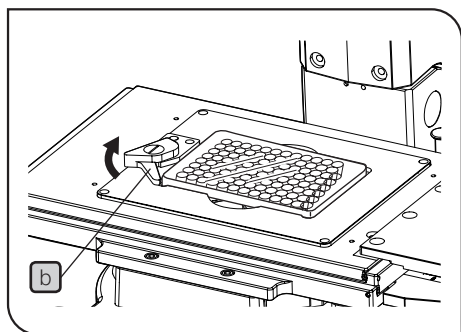
Open the specimen holder **b** of IX3-HOW, set the microplate in the center, push it toward the right diagonal direction, and return the specimen holder back to the original position.

«Mountable Microplate»

Microplate compliant with SLAS (ANSI/SBS Microplate Standards issued on Jan. 9, 2004.)

Size: 127.76 (plus or minus 0.5) x 85.48 (plus or minus 0.5) mm

Specimen holder: IX3-HOS, IX3-HO35D

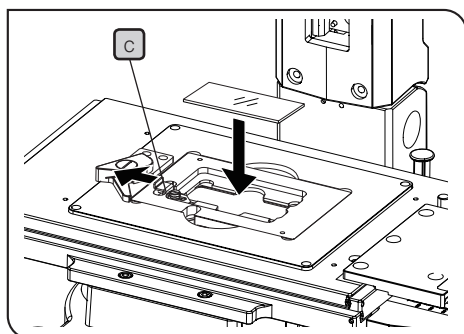


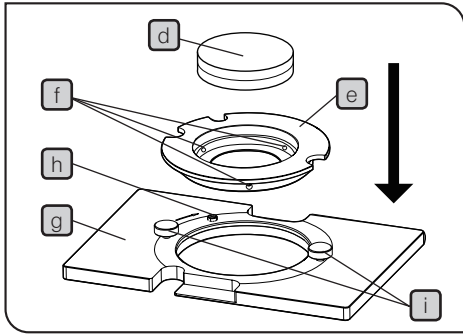
IX3-HOS

Open the specimen fixing part **c** of IX3-HOS outward, set the specimen in the center, push it toward the right diagonal direction, and return the specimen fixing part back to the original position.

«Chamber Slide Recommended»

- IWAKI Chamber Slide II (76 x 26 x 0.8 to 1.0 mm)
- Nunc Lab-Tek II Chamber Slide system (25 x 75 x 1.2 mm)
- BD Falcon CultureSlide (25 x 75 x 1.2 mm)

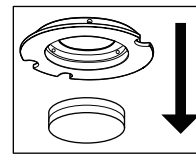




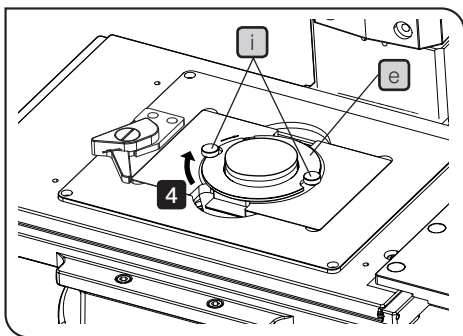
IX3-HO35D

- 1 Place the 35 mm dish **d** on the 35 mm dish fixing holder **e**. Tighten the fixing screws **f** (3 screws) placed on the side with the Allen screwdriver provided with IX3-HO35D to secure the 35 mm dish.

⊙ The 35 mm dish can be secured easily by tightening the fixing screws after flipping over the 35 mm dish in advance.



CAUTION Do not tighten the fixing screws too firmly. The dish may be damaged.



- 2 Set the fixing holder **e** in the center of IX3-HO35D **g** so that the cut-out meets the holder fixing knob **i**.
- 3 Loosen the holder fixing knobs **i**.
- 4 Rotate the fixing holder **e** clockwise to push it to the rotation stopper **h**.
- 5 Tighten the holder fixing knobs **i**.

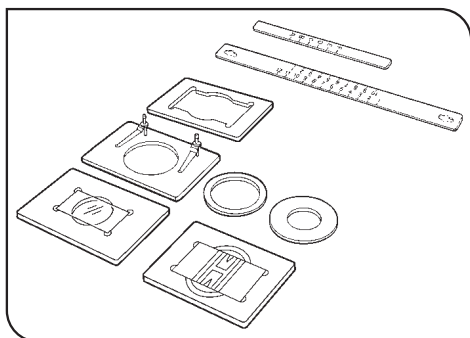
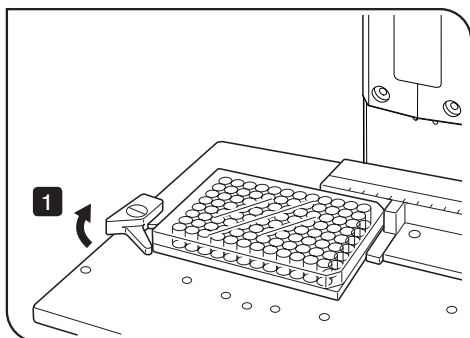
⊙ The 35 mm dish fixing holder **e** can be sterilized by using the autoclave.

«35 mm Glass Bottom Dish Recommended»

- Matsunami Glass D111310
- MatTek P35GC-1.5-14-C

«35 mm Dish Recommended»

- BD Falcon 351008



With the mechanical stage IX-MVR + stage IX2-SP

- ◎ 96-well or 24-well microplates, etc. are held in place by the specimen holder.

Microplates with dimensions of max. 136 mm x 92 mm can be accommodated in this way.

- 1** Open the spring-loaded finger of the specimen holder 1 and slide the microplate into the holder frame. Gently release the curved finger to clamp.

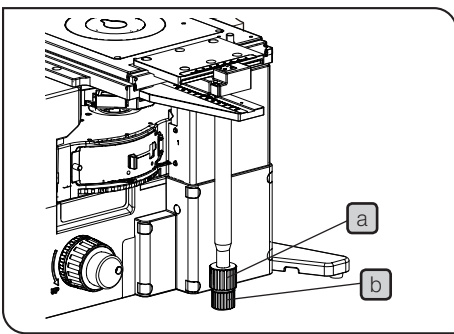
- ◎ To secure other vessels than microplates, various optional holders are available. A Terasaki plate holder is available for holding Terasaki plates (72-well, 60-well). When using this, it is necessary to replace the stage scales with those provided with the plate holder. Dish holders are available for 35 mm, 54 mm and 65 mm diameter dishes, a slide glass holder is available for holding slide glass, and the IX2-BCTP* is available for a blood cell test plate holder.

* A blood cell test plate or other calculating chamber for bacteria and eosinophil with mounting section dimensions corresponding to H 77 x V 35 x D 2 mm can be used. A 60 mm diameter dish can also be used.

2 Moving the Specimen

- CAUTION**
- Do not attempt to rotate the stage handle forcibly exceeding the stage movable range. The stage may be damaged.
 - As the objective may interfere with the stage depending on the focus position, be sure to operate carefully.

With the mechanical handle stage (IX3-SVR)



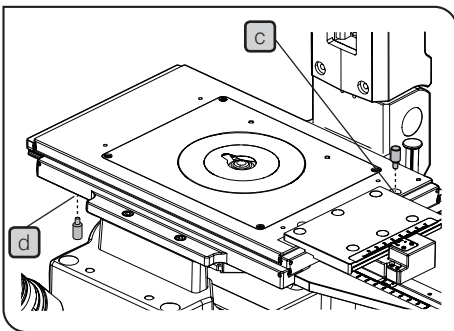
To move the specimen to a desired position, rotate the X-axis knob **a** and Y-axis knob **b**.

- ⊙ If the stage is used for a long time period, the stage movement range may be narrower rarely. In this case, move the stage several times within the full movement range toward the front/back or right/left direction while holding the top surface of the stage with both hands.

Movement control knob

Attaching the movement control knob provided with IX3-SVR makes it difficult to move the stage in the blocked direction. Even though you may touch the stage accidentally during observation, the observation position can be secured.

If the movement control knob is attached to the hole of **c**, the Y-axis movement will be blocked. If it is attached to the hole of **d** (backside of the stage), the X-axis movement will be blocked.



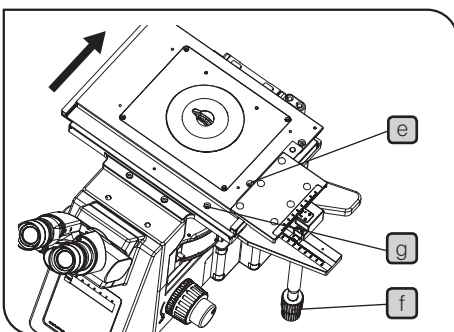
- CAUTION** Do not tighten the movement control knob too firmly. The stage may be damaged.

- ⊙ If you tighten the movement control knob while focusing on the specimen, it will be defocused.

Movement range limit screw

As a factory default, IX3-SVR, is equipped with the movement range limit screw which limits the stage movable range in the vertical or horizontal direction.

To enlarge the movable range to observe microplates, etc., remove the movement range limit screw.



- 1 Remove the movement range limit screw **e** in the vertical direction with the Allen screwdriver provided with the microscope.
- 2 Rotate the longitudinal handle **f** to move the stage inward.
- 3 Remove the movement range limit screw **g** in the horizontal direction with the Allen screwdriver provided with the microscope.

Stage Movable Range

With movement range limit screw :
50 mm in vertical direction, 50 mm in horizontal direction
(With the IX2-ILL30, IX3-ILL)

Without movement range limit screw:
70 mm in vertical direction, 114 mm in horizontal direction
(With the IX2-ILL30)
75 mm in vertical direction, 114 mm in horizontal direction
(With the IX3-ILL)

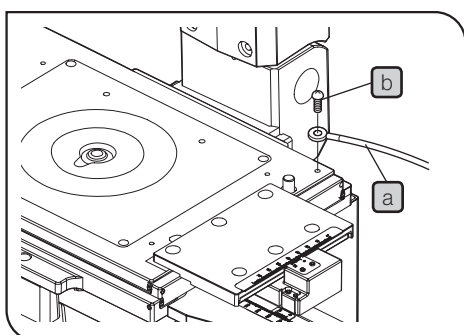
CAUTION When using the center plate provided with IX3-SVR, attach the movement range limit screw.

If the movement range limit screw is not attached, the objective may hit the stage.

With the Mechanical Stage IX-MVR

To move the specimen to a desired position, rotate the X-axis knob and Y-axis knob in the same manner as IX3-SVR.

◎ The stage travel area is 130 mm (X-axis) x 85 mm (Y-axis).



3 Connecting the Grounding Wire

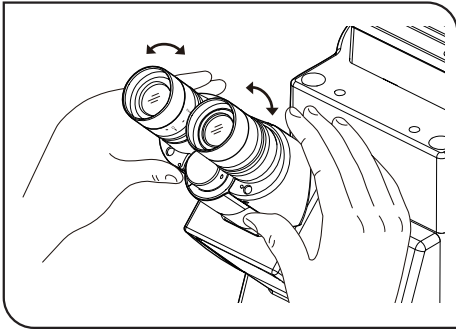
With the mechanical handle stage (IX3-SVR)

◎ A grounding wire can be attached to the stage for electrophysiological experiments, etc.

Prepare a grounding wire **a** and one M4 screw **b** and attach the grounding wire to a screw hole on the stage surface.

CAUTION The screw hole may sometimes be stuck by paint, etc. In such a case, screw in the M4 screw a few times to expose the metallic thread inside the screw hole and improve the contact before attaching the grounding wire firmly.

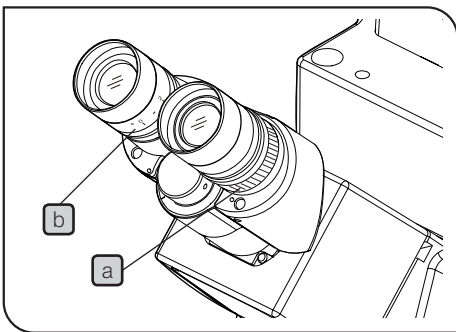
4-4 Observation Tube



1 Adjusting the Interpupillary Distance

While looking through the eyepieces, adjust the binocular vision until the left and right fields of view coincide completely. The index dot • indicates the interpupillary distance.

Ⓞ Note your interpupillary distance so that it can be quickly duplicated



2 Adjusting the Diopter

Ⓞ The diopter adjustment makes it possible to reduce the specimen focusing error even after the objective is switched. As the diopter varies between individuals, the diopter adjustment is required for each person.

CAUTION The eyepiece with diopter adjustment ring should always be inserted into the observation tube without the diopter adjustment ring.

- 1 Set the diopter adjustment rings on both sides to scale "0".
- 2 Engage a high-power objective (e.g. 40X) in the light path, look into the right eyepiece with your right eye, and bring the sample into focus using the coarse / fine adjustment knob.

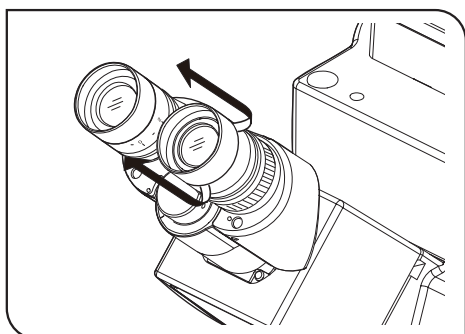
Ⓞ Do not use an immersion objective.

- 3 Engage a low-power objective (e.g. 10X) in the light path, rotate only the right diopter adjustment ring **a** to bring the sample into focus. At this time do not touch the coarse / fine adjustment knob.
- 4 Looking into the left eyepiece with your left eye, rotate only the left diopter adjustment ring **b** to bring the sample into focus.

Ⓞ The above procedure adjusts the diopter with reference to the right eye, but it is also possible to adjust with reference to the left eye. In this case, read the above procedure by inverting "right" and "left."

Using an eyepiece including a micrometer disk

- 1 Looking through the eyepiece with micrometer disk, turn the diopter adjustment ring **b** so that the micrometer in the field of view is sharply visible.
- 2 Looking through the eyepiece with micrometer disk, focus on the sample using the coarse / fine adjustment knob so that both the micrometer and sample are sharply visible.
- 3 Looking through the other eyepiece, turn only the diopter adjustment ring **a** to focus on the sample.



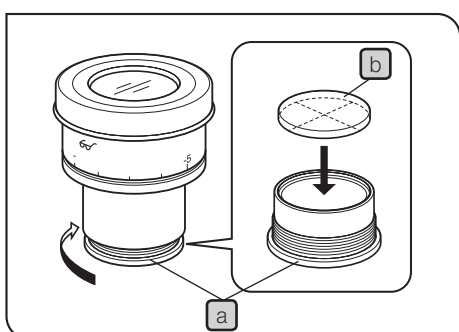
3 Using the Eye Shades

When wearing eyeglasses

Use the eye shades in the normal, folded-down position. This will prevent the eyeglasses from being scratched.

When not wearing eyeglasses

Extend the folded eye shades in the direction of the arrow to prevent extraneous light from entering between the eyepieces and eyes.



4 Mounting the Eyepiece Micrometer Disk

When the WHN10X-H eyepieces are used, an eyepiece micrometer disk can be mounted.

Use 24 mm dia. x 1.5 mm thick micrometer disks.

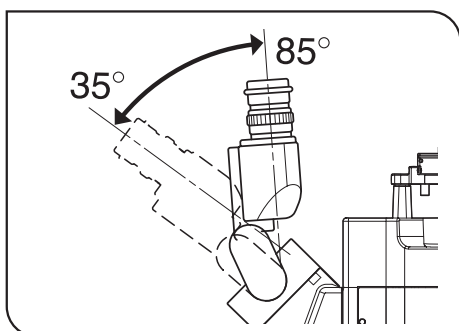
Turn the built-in micrometer-mounting frame **a** to the arrow direction (see figure) to remove it from the eyepiece and place a micrometer disk **b** into the mounting frame so that the surface with the model indication faces downward.

- ⊙ The micrometer-mounting frame may be too tight for certain micrometer disks.

In this case, turn the frame by holding the circumference with a light, uniform force or by applying the frame against a rubber sheet. Do not grasp the frame with a strong force, as this may deform the frame and make it harder to remove it.

Re-attach the micrometer mounting frame in the original position.

- ⊙ Be careful not to touch the lens or micrometer surface with your finger.



5 Adjusting the Tilt

- ⊙ Adjust the height and tilt of the eyepieces to obtain the most comfortable viewing position.

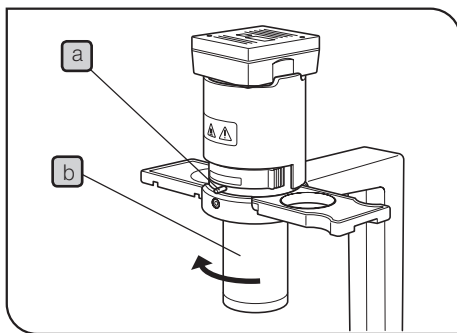
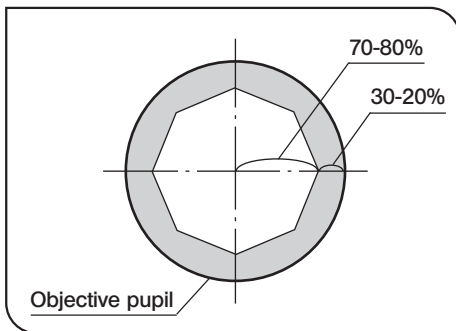
Holding the binocular section with both hands, adjust it to the desired position.

CAUTION

Never attempt to force the binocular section past the upper or lower stop position. Applying excessive force could destroy the limiting mechanism.

4-5 Illumination Column (IX2-ILL30)

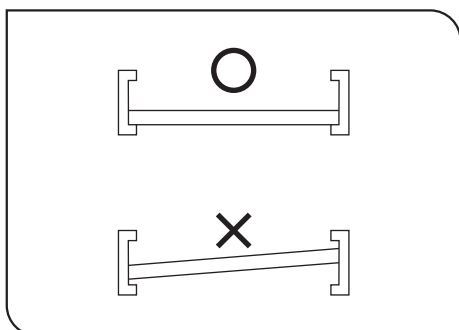
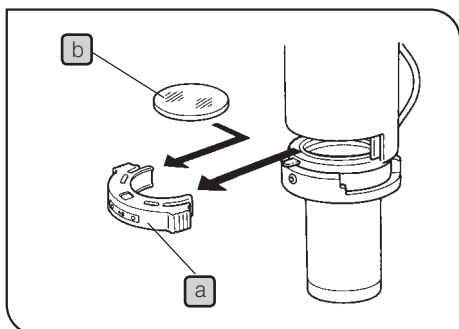
1 Using the Aperture Iris Diaphragm



- ⊙ The aperture iris diaphragm adjusts the numerical aperture of the illumination system in brightfield observation. It determines the focal depth, contrast and resolution according to purpose.
- ⊙ To check the aperture iris diaphragm, remove an eyepiece (and attach the U-CT30 if this is available) and look into the eyepiece sleeve. The aperture iris diaphragm can be seen as shown in left fig. Its aperture can be adjusted by rotating the aperture iris diaphragm lever **a**.
- ⊙ In general, the potential resolving power of an objective is fully utilized if the diaphragm is stopped down to correspond with the numerical aperture (N.A.) of the objective.

2 Removing the Condenser Lens

- ⊙ The working distance can be increased by removing the condenser **b** by rotating its bottom part in the clockwise direction when it is seen from above. However, the illumination becomes inappropriate. Use this method as a simplified method when using only large culture vessels.



3 Mounting the Filters

- ⊙ Wait until the filters have cooled down enough before replacing a filter. Take out the filter holder **a** and insert the required filter **b**.

CAUTION Insert the filter so that it comes at the bottom without tilting, as shown in left fig. If the filter is tilted or not inserted to the bottom, it may drop from the filter holder.

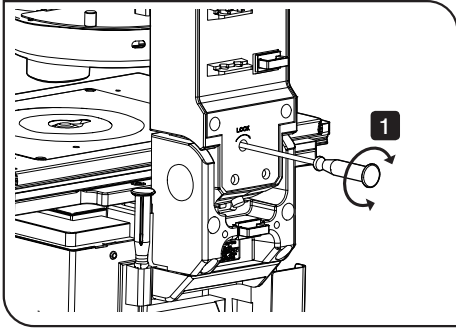
- ⊙ More than one filter can be stacked in the filter holder, provided that the total thickness of filters does not exceed 11 mm.

4 Using the Filters

- ⊙ Using appropriate filters according to the purposes allows you to observe and photograph specimens more effectively. Particularly, the use of the 45LBD filter is recommended in observation and photomicrography because it renders more neutral colors.
- ⊙ More than one filter with a diameter of 45 mm can be stacked in the filter holder **a**.

Filter	Application
43IF550-W45	Monochrome contrast filter (Green)
45ND6,45ND25	Light intensity adjustment filter (Transmittance 6% and 25%)
45LBD	Color temperature conversion filter (For observation and photomicrography)

4-6 Illumination Column (IX3-ILL)



1 Tilting the Illumination Column

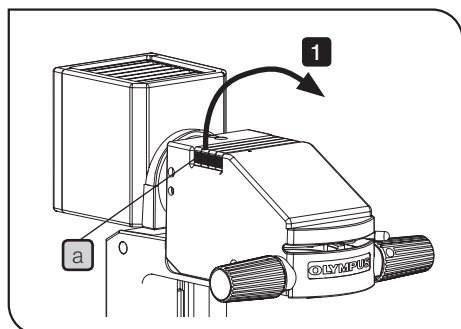
◎ When replacing large specimens, placing a micromanipulator or replacing a patch clamp electrode, working space can be created by tilting the illumination column.

1 Using the Allen screwdriver, turn the column tilt clamping screw to loosen it.

2 Holding the illumination column's upper side and slowly tilt the illumination column until it stops. Vibrations should be avoided. Accordingly, always support the illumination column with a hand and tilt slowly and gently. To return the column to its original position, reverse this procedure.

CAUTION

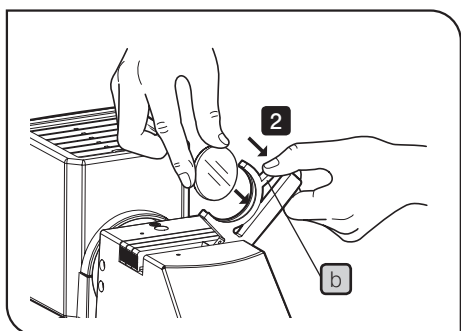
- When tilting the illumination column upwards or downwards, make sure not to catch your fingers in the hinge joint.
- The tilt clamping screw should normally be tightened during use. If the microscope is used while the screw is loosened, make sure that the illumination column does not accidentally tilt during use.
- Do not stop the illumination column while tilting. The illumination column may fall toward the front side or back side.
- When moving or transporting the microscope, always do so with the tilt clamping screw tightened.
- If a heavy module such as a high-intensity lamp housing is installed, always use the microscope with the tilt clamping screw tightened.
- Do not turn ON the illumination lamp while the illumination column is tilted.



2 Mounting Filters

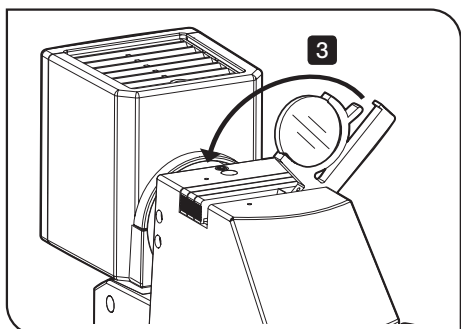
© 45 mm diameter, maximum 6 mm thick filters can be mounted. Various filters, such as the frost filter Ø45 mm (45FR), color temperature conversion filter (LBD), green interference filter (IF550) and ND filter can be mounted.

1 Place a finger on the milled section **a** of the filter holder and lift it.



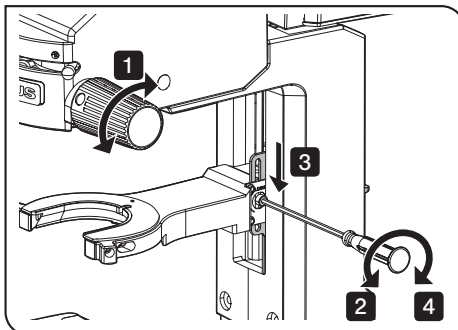
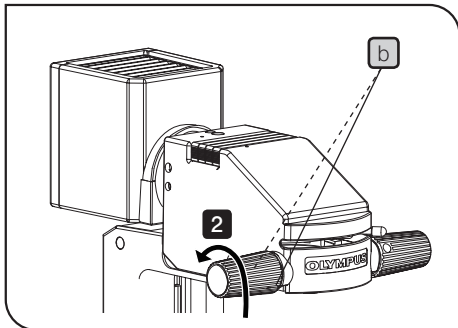
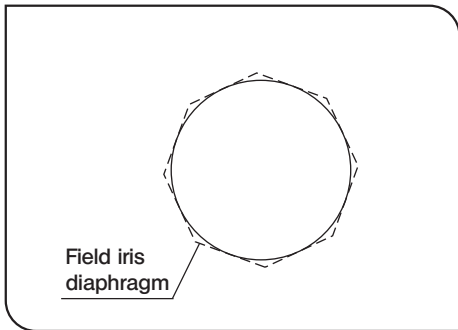
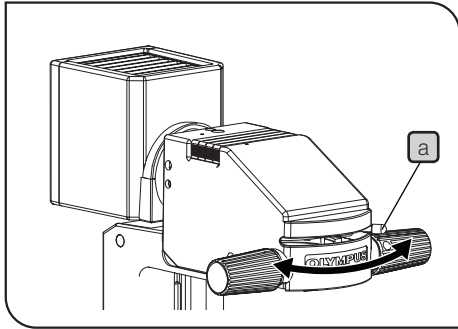
2 While holding the mounting lever **b** of the filter holder, insert a filter.

- CAUTION**
- Hold the filter by its edge to avoid leaving fingerprints or smudges on the filter surfaces.
 - Shortly after the transmitted light illumination has been used, the filter will be very hot. Be sure to set the main switch to "O" (OFF) and allow the filter holder and filters to cool down before replacing filters.



3 Return the filter holder.

- CAUTION**
- If the uneven brightness of the illumination light bothers you, we recommend engaging frost filter Ø45 mm (45FR), sold separately, in the light path.
 - If the filter holder is open, you may be dazzled by the light leaked from the light source in some cases. We recommend closing the filter holder before using the microscope



3 Using the Field Iris Diaphragm

⊙ The field iris diaphragm lever is used to adjust the diameter of the illumination beam in accordance with the objective in use. Adjust the diaphragm so that the field of view is circumscribed by the field iris diaphragm to cut extra light and improve the contrast of images.

- 1 Move the field iris diaphragm lever **a** to the left or right to close or open the diaphragm.

○ : Direction for opening the diaphragm

⊗ : Direction for closing the diaphragm

4 Adjusting the Condenser Height Adjustment Knob Tension

- 1 Loosen the two knob clamping screws **b** on the left adjustment knob using the Allen screwdriver.
- 2 While holding the right adjustment knob not to rotate it, turning the left adjustment knob counterclockwise (in the direction of the arrow) decreases the rotation tension and clockwise increases it. Rotating the right adjustment knob allows adjusting the tension of the condenser height adjustment knob while checking it.
- 3 After adjustment, tighten the two knob clamping screws **b** securely.

5 Condenser refocusing stopper

A mechanism returns the condenser back to the original position easily after moving the condenser.

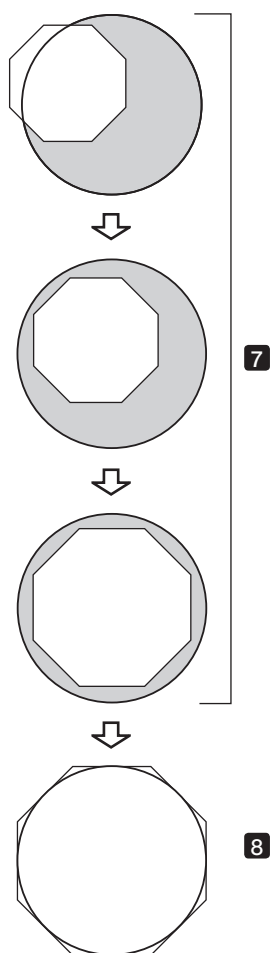
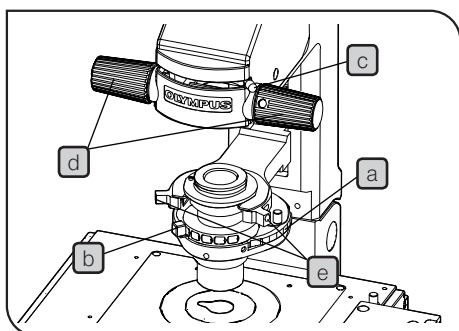
⊙ To use the condenser refocusing stopper, refer to page 47 , and attach the condenser refocusing stopper before use.

- 1 Bring the field diaphragm image into focus by rotating the condenser height adjustment knob.
- 2 Loosen the clamping screws of the stopper using the Allen screwdriver provided with the microscope.
- 3 Push the top of the stopper downward so that the stopper contacts the column securely.
- 4 Tighten the clamping screws of the stopper using the Allen screwdriver provided with the microscope.

CAUTION

Rotating the condenser height adjustment knob beyond the condenser height adjustment area with an excessive force could damage the microscope. Pay careful attention when rotating it.

⊙ If the manipulator is assembled to the column, the Allen screwdriver provided with the microscope may not be used in some cases.



6 Centering the Condenser

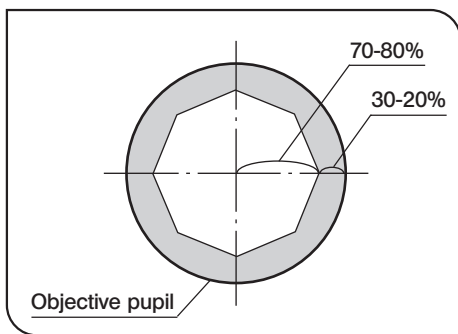
With the Condenser
(IX2-LWUCD, IX-ULWCD or IX2-MLWCD)

- 1 Rotate the turret **a** (either manually or electrically) to select the "BF" brightfield observation (with no optical element engaged in the light path).
- 2 Slide the aperture iris diaphragm lever **b** to open the diaphragm.
- 3 Slide the field iris diaphragm lever **c** to the fully open position.
- 4 Engage the 10X objective and bring the specimen into focus.
- 5 Using the field iris diaphragm lever, stop down the field iris diaphragm until its image is just inside the field of view.
- 6 Rotate the condenser height adjustment knob **d** to bring the field iris diaphragm image into focus.
- 7 While gradually opening the field iris diaphragm lever **c**, install the Allen screwdriver provided with the microscope in the adjustment hole **e** and rotate it so that the field iris diaphragm image is centered in the field of view of the eyepieces.
- 8 To check centration, open the field iris diaphragm until its image inscribes the field of view. Now the condenser is centered.

Effect of Field Iris Diaphragm

This is the iris diaphragm to adjust the area to be illuminated. By narrowing down to the level circumscribing the field of view depending on objectives, the excess light can be shielded to acquire the image with good contrast.

- ⊙ Depending on the condenser to be used, the field iris diaphragm may not be viewed by the objective of 40X or higher magnification.



7 Using the Aperture Iris Diaphragm

- ⊙ In general, the potential resolving power of an objective is fully utilized if the diaphragm is stopped down to correspond with the numerical aperture (NA) of the objective.
- ⊙ Depending on the specimen, image contrast or focal depth in observation or acquisition may be improved by keeping the aperture iris diaphragm stopped down a little. In general, a good image is obtained if the diaphragm is stopped down to between 70% and 80% of the NA of the objective. Stop further down for less contrasty specimens.
- ⊙ To check the position of the perimeter of the aperture iris diaphragm, remove the eyepieces and look into the eyepiece sleeves to view the aperture iris diaphragm image and the objective's exit pupil.

4-7 Oil- or Water-Immersion Objective

1 Using Oil- or Water-Immersion Objective

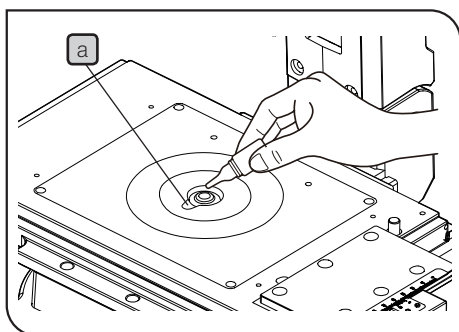
☉ If you use an oil-immersion objective, use immersion oil as described below.

CAUTION Always use immersion oil supplied by Olympus.

1 Using a low-power objective, bring the specimen into focus.

2 Rotate the revolving nosepiece to engage the oil immersion objective.

3 Remove the specimen and move the stage insert cut-out **a** close to the objective front lens. Apply a drop of the immersion oil to the objective front lens. Place the specimen and rotate the fine adjustment knob to bring the specimen into focus.



CAUTION · Use as little oil as possible.

· If the oil contains air bubbles, the image will be degraded. Make sure the oil is free of air bubbles.

4 After use, remove immersion oil from the objective front lens by wiping with gauze slightly moistened with absolute alcohol.

☉ The presence of air bubbles can be checked by viewing the pupil of the objective (viewed as a bright circular shape) in the tube after removing the eyepiece and opening the field iris diaphragm and the aperture iris diaphragm completely.

CAUTION Caution in use of immersion oil:

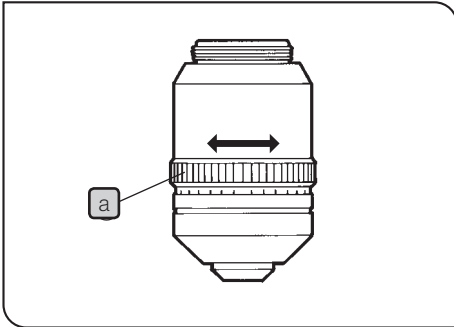
If immersion oil enters your eyes or contacts your skin, immediately take the following treatment.

Eyes: Rinse with fresh water (for 15 minutes or more).

Skin : Rinse with water and soap.

If the appearance of the eyes or skin is altered or pain persists, immediately see your doctor.

4-8 Objectives with Correction Collar



⊙ If the thickness of the cover glass or the vessel does not match the thickness scale of an objective with correction collar, the objective cannot manifest its performance. When using a correction collar equipped objective, perform the following adjustment as required.

Adjustment Procedure

- If the cover glass thickness is known, set the correction collar **a** to that value on the scale.
 - If the cover glass thickness is unknown, adjust the correction collar **a** and rotate the fine adjustment knob alternately until the positioning with the highest contrast is obtained.
- ⊙ Be careful not to touch the correction collar **a** when turning the revolving nosepiece.

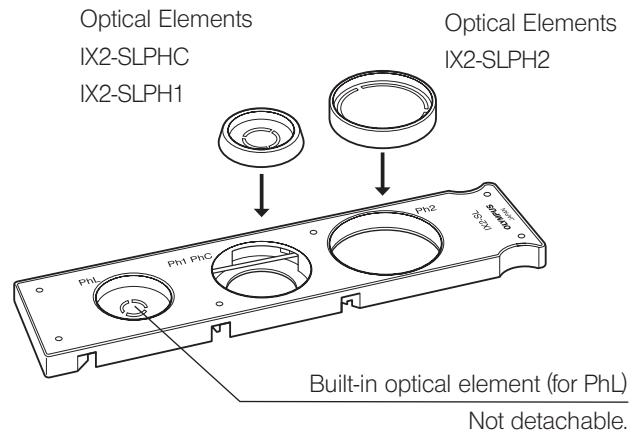
5 OTHER OBSERVATION METHODS

5-1 Phase Contrast Observation

© A phase contrast objective, phase contrast optical element, and the U-CT30-2 centering telescope are required for phase contrast observation.

1 Phase Contrast Optical Elements and Applicable Objectives

Phase Contrast Slider
IX2-SL



Optical Element	Applicable Objectives
PHL (built-in)	UPLFLN4XPH
IX2-SLPH1	CPLN10XPH, LCACHN20XPH, CPLFLN10XPH
IX2-SLPHC	UPLFLN10X2PH, UPLFLN20XPH, LUCPLFLN20XPH
IX2-SLPH2	UCPLFLN20XPH, UPLFLN40XPH, LUCPLFLN40XPH, LUCPLFLN60XPH, LCACHN40XPH

With the or IX2-LWUCD

- ◎ Insert the optical element (small) in the 30 mm position and the optical element (large) in the 38 mm position. When observing the specimens in wells, it is recommended to use the IX-PHC to obtain the phase contrast effect in a wide range of field of view.

Optical Element	Indication	Applicable Objectives
IX-PHL (small)	PhL	UPLFLN4XPH
IX-PHC (small)	PhC	CPLN10XPH, LCACHN20XPH, CPLFLN10XPH
IX-PH1 (small)	Ph1	UPLFLN10X2PH, UPLFLN20XPH, LUCPLFLN20XPH
IX-PH2 (small)	Ph2	UCPLFLN20XPH, UPLFLN40XPH, LUCPLFLN40XPH, LUCPLFLN60XPH, LCACHN40XPH
IX-PH3 (large)	Ph3	PLAPON60XOPH, UPLFLN60XOIPH, UPLSAPO100XOPH, UPLFLN100XO2PH

With the IX-ULWCD

- ◎ The IX-PHCU or IX-PH1U can be attached only in the Ph1 and PhC. (Do not remove the built-in elements.)

Optical Element	Indication	Applicable Objectives
PHL (built-in)	PhL	UPLFLN4XPH
IX-PHCU	PhC	CPLN10XPH, LCACHN20XPH, CPLFLN10XPH
IX-PH1U	Ph1	UPLFLN10XPH, UPLFLN20XPH, LUCPLFLN20XPH,
PH2 (built-in)	Ph2	UCPLFLN20XPH, UPLFLN40XPH, LUCPLFLN40XPH, LUCPLFLN60XPH, LCACHN40XPH

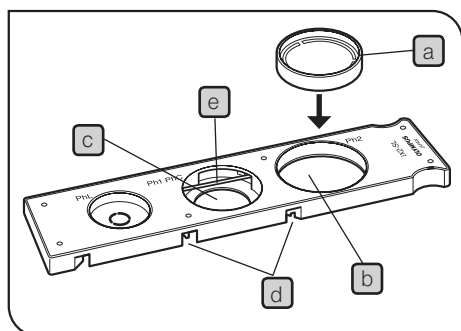
- ◎ When using the U-UCD8 or IX2-MLWCD, refer to the provided instructions.

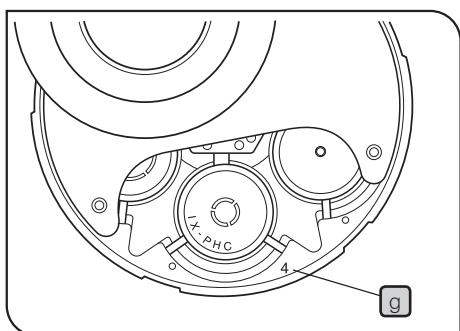
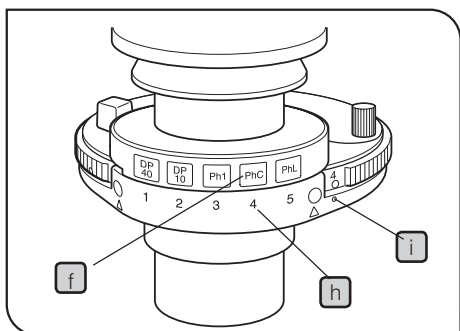
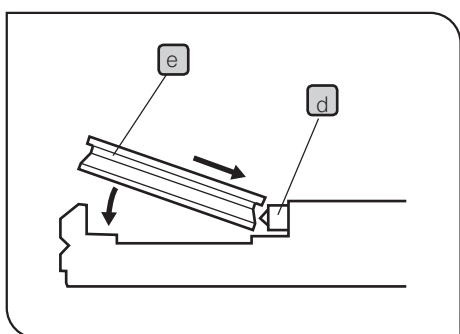
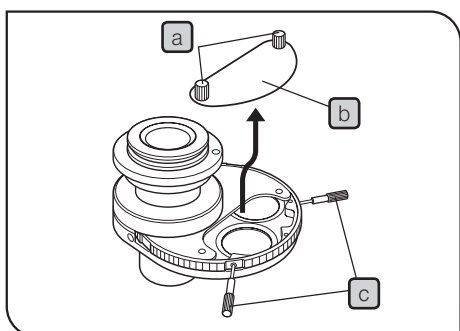
2 Attaching the Optical Elements

IX2-ILL30

- ◎ When using an objective that is not compatible with the built-in optical element, it is required to use a special optical element.

- 1 To insert the IX2-SLPH2 optical element, insert it in the idle position **b** so that the surface with indication **a** faces upwards.
- 2 When attaching the optical element IX2-SLPHC or IX2-SLPH1 in the center position **c**, insert the provided centering knobs into the screw holes **d**, loosen them, insert the optical element passing under the line spring **e** and tighten lightly the centering knobs again.





IX3-ILL(With the IX2-LWUCD, IX-ULWCD)

⊙ Do not engage any optical element in the BF (brightfield) light path.

- 1 Place the condenser in a direction of left-hand figure, loosen the detaching screw **a** and remove the cover **b**.
- 2 Rotate the turret so that the number of the next optical element to be inserted in the uncovered position is visible.
- 3 Loosen the optical element position centering screws using the optical element centering knobs **c**.
- 4 Hold a phase contrast ring and, while pushing the spring **d** inside the turret with the edge of the phase contrast ring **e**, insert the ring completely in the turret position until the ring frame contacts the bottom of the position.
- 5 Rotate the optical element centering knobs clockwise to tighten the centering screws lightly.
- 6 Place the index **f** provided with each optical element in the index insertion hole having the same number **h** as the number of the position **g** in which the corresponding optical element is mounted. (IX-ULWCD does not have the index.)

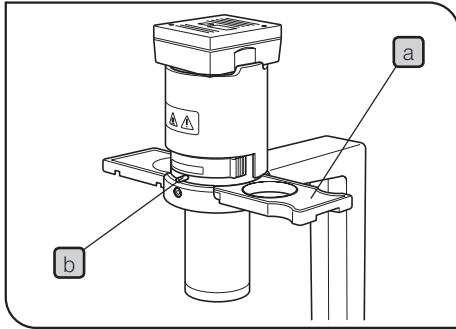
⊙ The optical element with the number indicated by marking **i** is engaged in the light path.

- 7 When all of the required optical elements have been mounted, attach the cover and tighten the detaching screws.

- CAUTION**
- Be careful not to apply pressure to the ring slit inside the frame.
 - If the optical element centering knobs are attached, the turret is unable to be rotated.
 - If the optical element fixing screws are loosened, the fixing screws will interfere while rotating the turret. Be sure to tighten the screws firmly. Do not tighten the optical element centering knobs too much, this may deform the frames of the optical elements.
 - To remove an optical element index, use the tip of a ball-point pen or mechanical pencil.

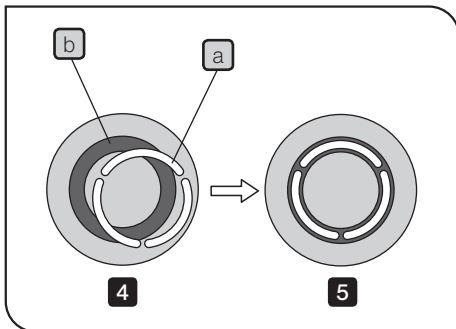
With the IX2-MLWUCD

⊙ Refer to the provided instructions.



3 Attaching the Phase Contrast Slider

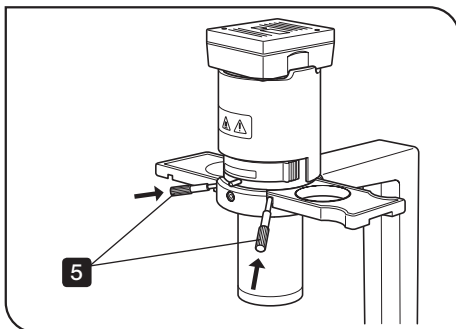
- 1 Attach the phase contrast slider **a** to the illumination column so that the slider's indication surface faces upwards and the finger hook position comes on the right.
- 2 Be sure to set the aperture iris diaphragm lever **b** to the open position (O) for phase contrast observation.



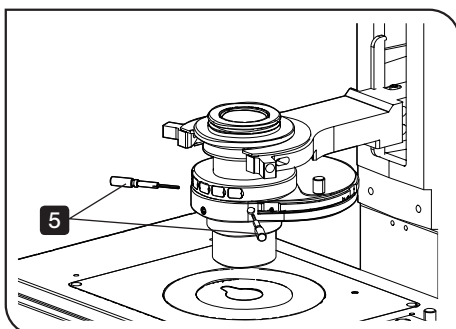
4 Centering the Phase Contrast Ring Slit

- ⊙ Open the aperture iris diaphragm during phase contrast observation.
- ⊙ The IX2-SLPH2 optical element does not need centering.

- 1 Engage the phase contrast objective in the light path and bring the specimen into focus.
- 2 Remove an eyepiece and attach the U-CT30-2 centering telescope in place.
- 3 Engage the ring slit of the condenser matching the phase contrast objective in the light path.
- 4 Rotate the knurled section of the centering telescope to focus on the ring slit **a** and the phase plate **b** of the objective.
- 5 Using the optical element centering knobs, turn the phase contrast ring slit centering screws (in positions marked) so that the ring slit image overlaps with the phase plate of the objective.
- 6 Remove the U-CT30-2 centering telescope and attach an eyepiece in place.



IX2-ILL30



IX3-ILL

- ⊙ If the vessel is not completely flat, it may become necessary to adjust the centering again to obtain the optimum contrast. Adjust the centering in each objective power.
- 7 Adjust the field iris diaphragm so that its image circumscribes the field of view and observe the phase contrast.
- ⊙ Engaging the green filter in the light path will improve the contrast.

5-2 Reflected Light Fluorescence Observation (Separate Manual)

© Refer to the separate instruction manual (REFLECTED FLUORESCENCE SYSTEM).

6 CAMERA RECORDING

1 Camera Adapter

The observed images can be acquired by attaching the camera adapter and the microscope digital camera to the left side port of IX53.

- ⊙ Be sure to adjust the parfocality before using a camera adapter. Otherwise, the focusing of the camera image will not match that of the image observed through eyepieces. When attaching the camera adapter, refer to the instructions of the camera adapter in use, too. For parfocality adjustment procedures, refer to the Instruction Manuals of the Camera Adapters.
- ⊙ When attaching the camera to the left side port, it is recommended to mount the support (column) between the camera and table top surface. If the camera is tilted, decentering may occur while replacing the objective lens.

2 Division of Light Paths

The light paths are divided as follows:

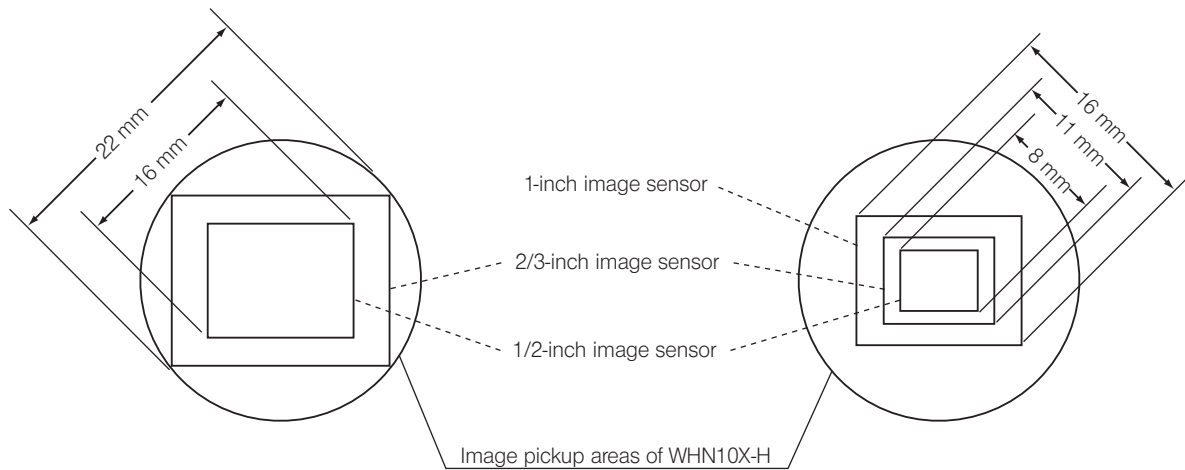
Left side port 50% light path / Observation 50% light path

- ⊙ The light such as the fluorescent lamp may enter from the eyepiece to be reflected in the acquired image.

3 Selecting the Camera Adapter Magnification

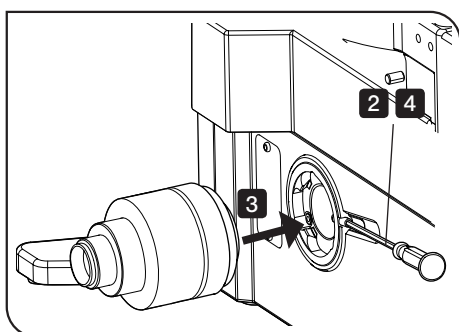
The image pickup area is determined by the size of the image sensor used in the TV camera or digital camera and the magnification of the camera adapter.

The following illustrations show the image pickup areas of camera adapters when using the WHN10X-H eyepieces (FN 22).



When a 0.5X camera adapter is used

When a 1X camera adapter is used



4 Installing the Camera Adapter

⊙ When attaching the camera adapter, refer to the instructions of the camera adapter in use, too.

- 1** Screw the camera adapter securely into the C-mount camera.
- 2** Using the Allen screwdriver, loosen the left side port clamping screw of the microscope to remove the cap.
- 3** Insert the mount dovetail of the camera adapter into the camera adapter mount of the trinocular tube.
- 4** Tighten the clamping screw firmly.

⊙ When the left side port is not used, attach the cap.

⊙ If U-DPCAD is attached to the left side port, the camera cannot be attached to the inside camera port depending on the camera size.

⊙ To acquire the 1X image by the left side port, combine U-TV1X-2 with U-CMAD3.

⊙ Attaching a heavy camera to the left side port may allow decentering the image due to the tilted adapter. It is recommended to reinforce the installation using a commercially available lab jack, etc.

7 TROUBLESHOOTING GUIDE

Under certain conditions, performance of the microscope may be adversely affected by factors other than defects. If problems occur, please review the following list and take remedial action as needed.

If you cannot solve the problem after checking the entire list, please contact your local Olympus representative for assistance.

Problem	Cause	Remedy	Page
a) The bulb does not light.	Power cord of the TL4 is unplugged.	Plug the power cord into a power outlet.	49
	Main switch of the TL4 is not "I" ON.	Set the main switch to "I" (ON).	13
	Bulb is burnt out.	Replace the bulb.	43
b) The bulb lights but the field of view is dark.	Lamp voltage is too low.	Increase the light intensity to an optimum voltage.	13
	Condenser is not well positioned.	Adjust the condenser height until the field iris diaphragm image is formed in the specimen plane.	27
	Too many filters are used.	Reduce the number of filters to the minimum required.	25
	Stage center plate is engaged in the light path.	Move the stage and place the specimen again.	15
	Field iris diaphragm is not opened wide enough.	Open the field iris diaphragm sufficiently.	26
c) Field of view is obscured or not evenly illuminated.	An objective that falls outside the condenser's illumination range is used.	Use a condenser that matches the objective.	27
	Field iris diaphragm is not properly centered.	Center the field iris diaphragm correctly.	27
	Field iris diaphragm is stopped down too far.	Open the field iris diaphragm sufficiently.	26
	A filter is stopped in an intermediate position.	Set the filter at the appropriate position.	25
d) Dirt or dust is visible in the field of view.	Dirt/dust on the specimen.	Clean thoroughly.	5
	Dirt/dust on the eyepieces.		
	Dirt/dust on a mirror unit.		
	Dirt/dust on the optical element.		
	Condenser is not correctly positioned and the frost filter Ø45 mm (45FR) or filter is focused.	Adjust the condenser height until the field iris diaphragm image is formed in the specimen plane.	27
e) Image glares.	Condenser is raised too high.	Lower to the proper position.	27
	Aperture iris diaphragm is stopped down too far.	Open the aperture iris diaphragm.	28

Problem	Cause	Remedy	Page
f) Visibility of observed image is poor. <ul style="list-style-type: none"> • Image is not sharp. • Contrast is poor. • Details are poorly visible. 	Objective in use is not designed for UIS2 series.	Replace with an objective designed for UIS2 optics.	-
	Correction collar on the objective equipped with correction collar is not adjusted.	Adjust the correction collar to acquire the best contrast.	30
	Front lens of the objective is dirty.	Clean the objective.	5
	The immersion oil appropriate with an oil immersion objective is not used.	Use Olympus immersion oil with the oil immersion objective, Olympus silicone oil with the silicone immersion objective and water with the water immersion objective.	29
	Immersion oil contains bubbles.	Remove bubbles.	-
	Inappropriate slide or cover glass thickness.	Replace with glass of appropriate thickness.	-
	Glass components (condenser, objective, eyepieces, culture vessels, etc.) are dirty.	Clean thoroughly.	5
	Ring slit and phase plate are not centered.	Center them correctly.	34
g) A part or one side of the field of view is blurred.	Specimen is tilted with respect to the stage.	Place the specimen correctly on the stage and secure it with the stage clip.	15
h) Field of view of one eye does not match that of the other.	The interpupillary distance is incorrect.	Adjust the interpupillary distance.	20
	Incorrect diopter adjustment.	Adjust the diopter.	20
	You are not accustomed to parallel optical axis.	When looking into the eyepieces, do not stare at image from the beginning but see the overall field of view. It is sometimes recommended to turn your eyes away from the eyepieces, look far off and look into the eyepieces again.	-

8 SPECIFICATIONS

Item	Specification		
Optical system	UIS2 optical system		
Microscope Frame IX53P1F	Division of light path 50% for observation light path, 50% for left side port Focusing movable range Upper side: 6.5 mm or more from the original position Lower side: 3 mm or more from the original position Original position: 1 mm above the stage surface Left side port image magnification: 1X		
Illumination Column	Type	IX2-ILL30	IX3-ILL
	Movement mechanism	Column for installing the U-LS30-3 lamp socket. Provided with the condenser (NA 0.3, WD 72 mm) Designated bulb 6 V, 30 W halogen bulb 6 V 30 WHAL (PHILIPS 5761) Power supply: Halogen lamp power supply unit TL4	Column for installing the lamp housing, with 30° tilting mechanism. Condenser holder up/down movement range: 88 mm Condenser refocusing mechanism is available. Designated bulb 12 V, 100 W long-life halogen bulb 12V100WHAL-L (PHILIPS 7724) Power supply: Halogen lamp power supply unit TH4
Observation Tube	Type	U-BI90	U-TBI90
		Binocular	Tilting binocular
	Field No.	22	
	Tube inclination	45°	35° to 85°
	Interpupillary distance adjustment	50 to 76 mm	
Diopter adjustment range	- 5 to + 5 diop		
Stage	Type	IX3-SVR	IX2-SP
	Size	240 mm (D) x 444.5 mm (W)	240 mm (D) x 232 mm (W)
	Movement mechanism	<ul style="list-style-type: none"> · X- and Y-axis knob with adjustable tension. · Right long axis handle (can be mounted by reversing the right and left) 	Not available
		Movement range: (Combined with IX2-ILL30): 70 mm vertical (Y), 114 mm horizontal (X). (Combined with IX3-ILL): 75 mm vertical (Y), 114 mm horizontal (X).	Movement range (Combined with IX-MVR): 85 mm vertical (Y), 130 mm horizontal (X).
Option	Replaceable center plate (Ø110 mm)	Replaceable center plate (Ø110 mm)	
Operating environment	<ul style="list-style-type: none"> • Indoor use. • Altitude: Max. 2000 meters • Ambient temperature: 5 to 40 °C (41 to 104 °F) • Maximum relative humidity: 80 % for temperatures up to 31 °C (88 °F) (without condensation) In case of over 31 °C (88 °F), the relative humidity is decreased linearly through 70 % at 34 °C (93 °F), 60 % at 37 °C (99 °F), and to 50 % at 40 °C (104 °F). • Supply voltage fluctuations: ±10% • Pollution degree: 2 (in accordance with IEC60664-1) • Installation/Overvoltage category: II (in accordance with IEC60664-1) 		
Transportation/ storage environment	<ul style="list-style-type: none"> • Temperature: Min. -25°C, Max. 65°C • Humidity: Min. 0%, Max. 90% (without condensation) 		


9 ASSEMBLY

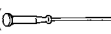

To ensure the performance, we recommend that you have your Olympus representative to assemble this microscope.

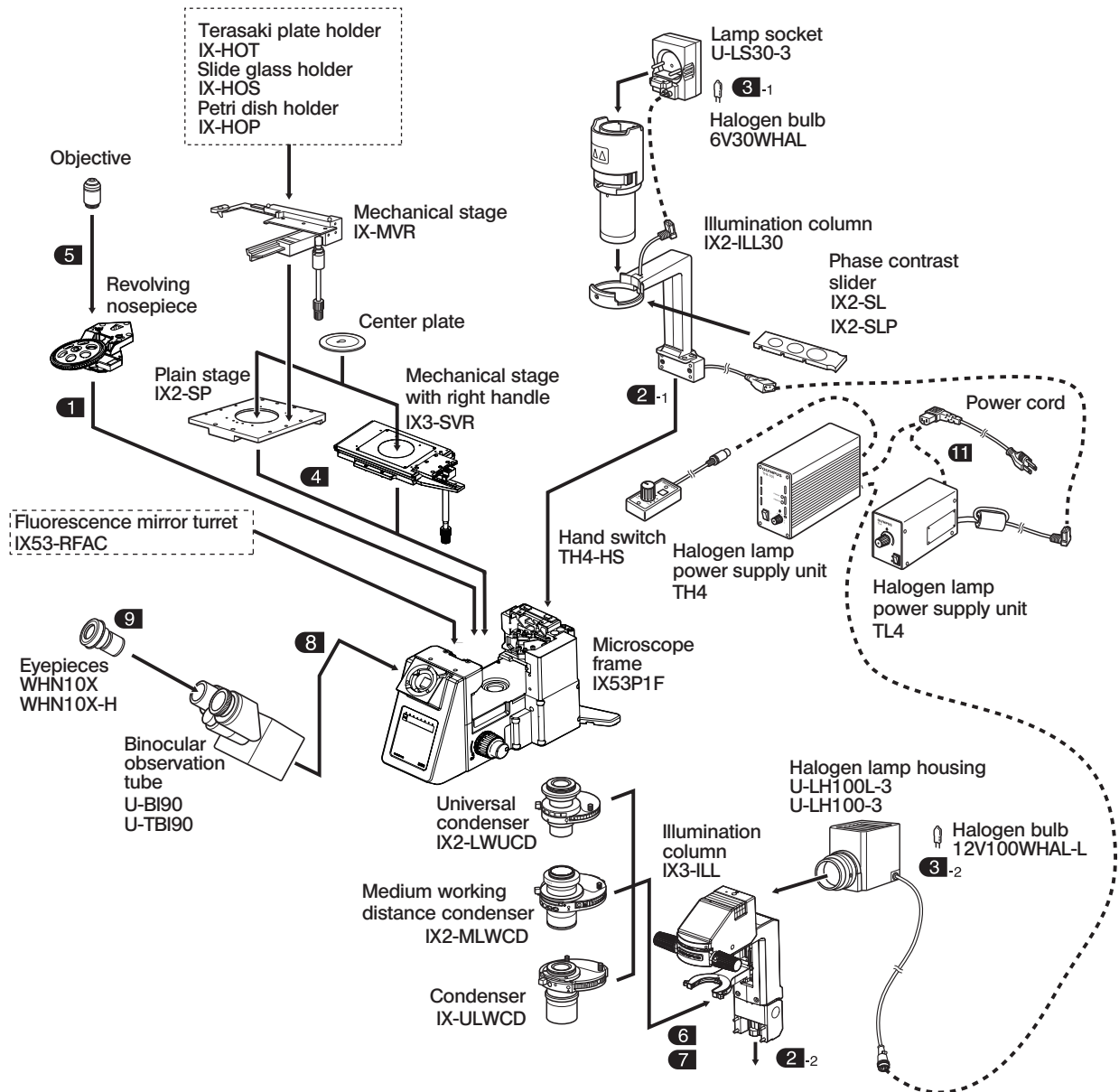
9-1 Assembly Diagram

The diagram below shows the sequence of assembly of the modules. The numbers indicate the order of assembly. The modules shown in the following diagram are merely the basic ones. For the modules which are not shown in the diagram, please consult your Olympus representative or the latest brochures.

When assembling the microscope, make sure that all parts are free of dust and dirt, and avoid scratching any parts or touching glass surfaces. Also be sure to release the transport lock of the revolving nosepiece and light path selector by removing the screws before use (see pages 1 & 2).

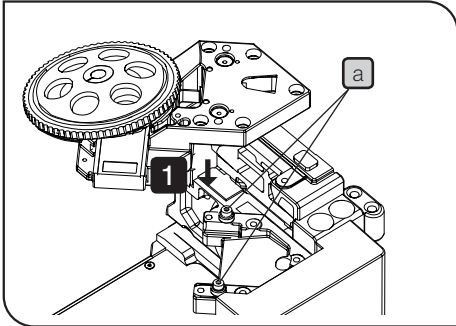
Assembly steps enclosed in  will be detailed on the subsequent pages.

© All assembly operations are possible by using the 3 mm Allen screwdriver () and 4 mm Allen wrench () provided with the microscope. For assembly of the reflected fluorescence system and modules without descriptions, refer to the separate instruction manuals.



9-2 Detailed Assembly Procedures

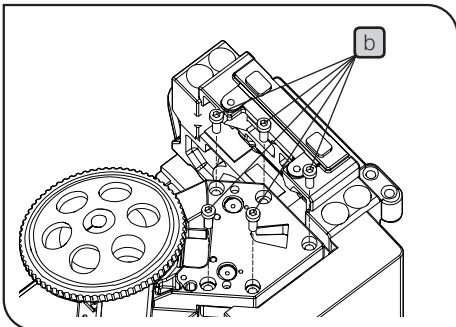
1 Attaching the revolving nosepiece



CAUTION To ensure the performance, we recommend that you have your Olympus representative to assemble the revolving nosepiece.

- 1 Insert the revolving nosepiece slowly into the positioning pins **a** (2 positions) in the upper area of the microscope.

CAUTION As the mounting holes of the positioning pins are small, insert the revolving nosepiece into the positioning pins carefully from the above paying attention not to tilt the revolving nosepiece.

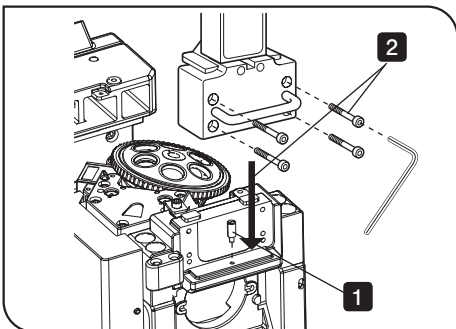


- 2 Tighten the revolving nosepiece fixing screws **b** (5 positions) by using the Allen wrench provided with the microscope.

2 Mounting the Illumination Column

CAUTION Do not tighten the screws with the illumination column inclined. Doing so may damage the screw sections.

IX2-ILL30

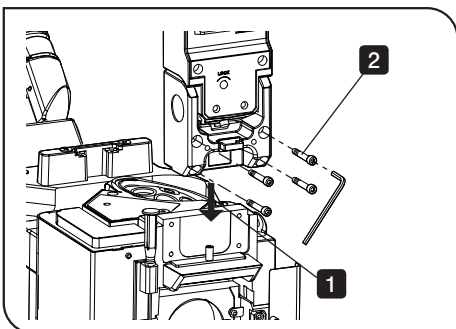


- 1 Loosen the protruding positioning pin on the microscope frame to remove, by using a minus driver.

- 2 While holding the illumination column with one hand, insert the four provided Allen screws into the screw holes. Then tighten the screws with the provided Allen wrench.

CAUTION Hold the illumination column securely with your hand until the screw is tightened. As a positioning pin is removed, the illumination column may be fallen from the microscope frame.

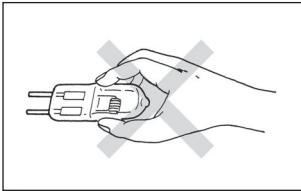
IX3-ILL



- 1 Aligning the two guide holes on the illumination column with the two protruding positioning pins on the microscope frame, gently fit the column onto the microscope frame from the above.

- 2 While holding the illumination column with one hand, insert the four provided Allen screws into the screw holes. Then tighten the screws with the provided Allen wrench.

3 Attaching the Halogen Bulb



CAUTION To prevent reduced bulb life or cracking, do not touch the bulb with bare hands. If fingerprints are accidentally left on the bulb, wipe the bulb with a dry soft cloth.

Caution for Bulb Replacement During or Right After Use

The bulb, lamp housing and areas near these will be extremely hot during and right after use.

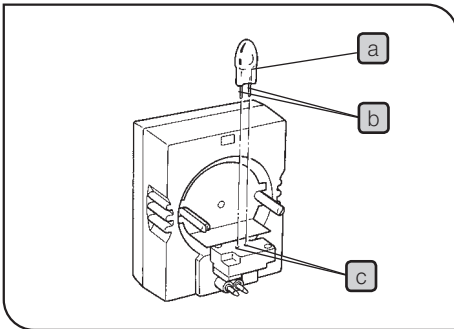
Set the main switch to "O" (OFF), disconnect the power cord from the wall outlet, then allow the old bulb and lamp housing to cool before replacing the bulb with a new of the designated type.

IX2-ILL30

Ⓢ The designated bulb model is the 6V30W/30V (PHILIPS 5761) halogen bulb.

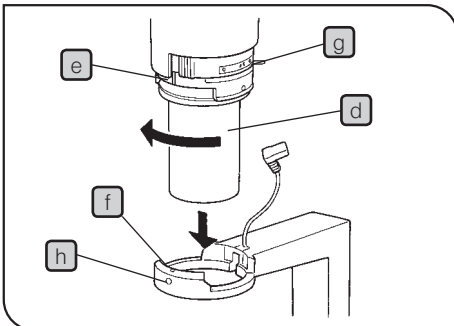
- 1 Hold the halogen bulb **a** with gloves or a piece of gauze, insert the bulb pins **b** straight and fully into the pin holes **c** on the lamp socket.

CAUTION Push in gently. If an excessive force is applied or the bulb is twisted, the bulb may be damaged.



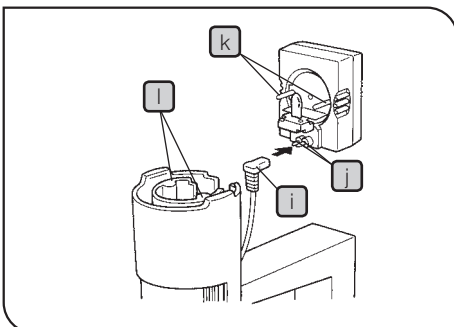
Attaching the Condenser

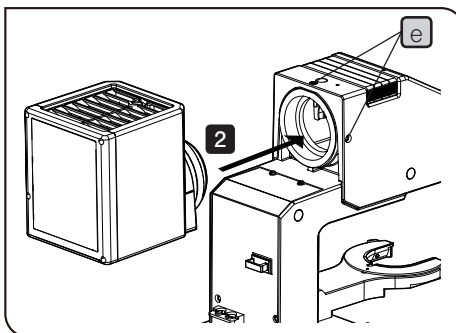
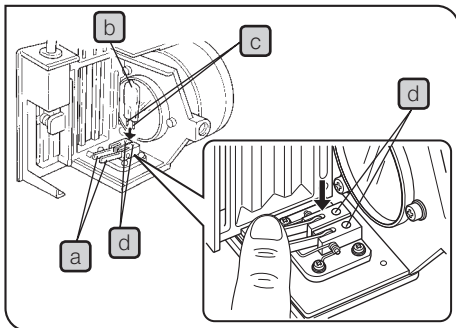
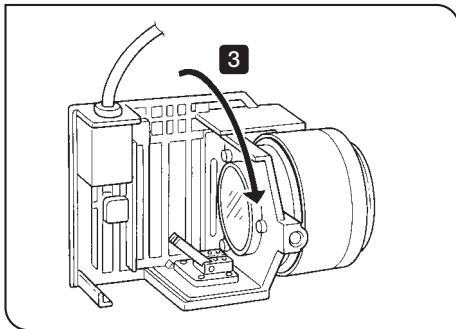
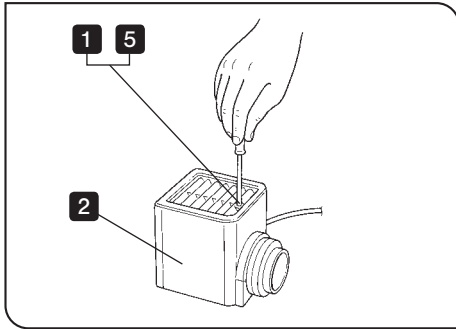
- 1 Align the index groove **e** on the condenser **d** with the dent section **f** of the illumination column and fit the condenser **d**.
- 2 Rotate the condenser **d** by 90° clockwise so that characters "AS" **g** on the filter holder surface faces the front, then attach and tighten firmly the provided clamping screw **h** using the Allen wrench provided with the microscope frame.



Attaching the Lamp socket

Ⓢ Insert the plug **i** into the socket **j**, then push the guide pins **k** gently into the guide holes **l** on the condenser.





IX3-ILL

© The designated bulb model is the 12V100WHAL-L (PHILIPS 7724) halogen bulb.

1 Fully loosen the clamping screw at the top of the halogen lamp housing using the Allen screwdriver provided with the microscope frame.

2 Lift up the halogen lamp housing to remove it.

3 Tilt the bulb socket by 90 ° in the direction of the arrow.

4 While pushing down the bulb clamping lever **a**, hold the halogen bulb **b** with gloves or a piece of gauze, insert the bulb pins **c** straight and fully into the pin position **d** on the lamp socket. Then return the lamp clamping lever gently back to the original position to clamp the bulb.

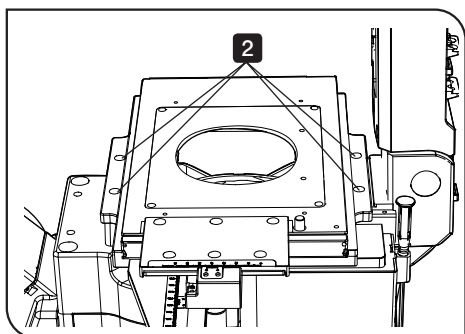
5 Fit the halogen lamp housing from up and tighten the clamping screw by applying downward pressure.

Mounting the Lamp Housing

1 Loosen the two clamping screws **e** of the illumination column with the Allen screwdriver provided with the microscope.

2 Fit the lamp housing to the illumination column

3 Tighten the two clamping screws **e** of the illumination column.



4 Attaching the Stage

⊙ The following stage mounting procedures are commonly applicable to the IX3-SVR, IX2-SP.

⊙ When mounting the mechanical stage IX-MVR, attach first the plain stage IX2-SP in advance (see the description on the bottom of this page).

1 Gently place the stage on the microscope frame by aligning the stage mounting holes with the threaded holes on the frame.

2 Insert the four provided Allen screws into the mounting holes. Tighten the screws using the provided Allen wrench.

⊙ Mount the stage so that the vertical (horizontal) direction of the stage is parallel to the vertical (horizontal) direction of the microscope.

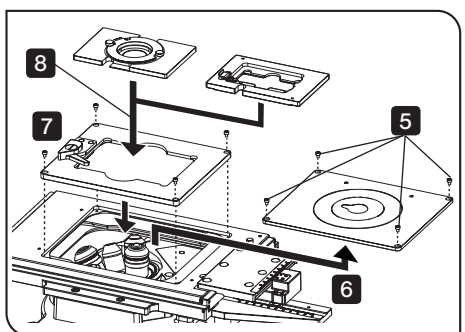
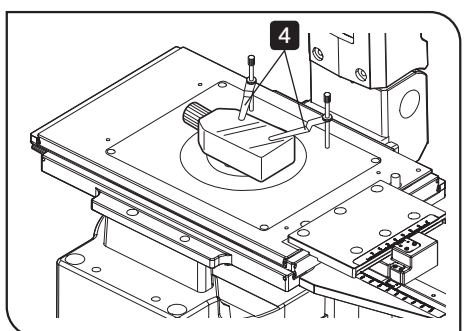
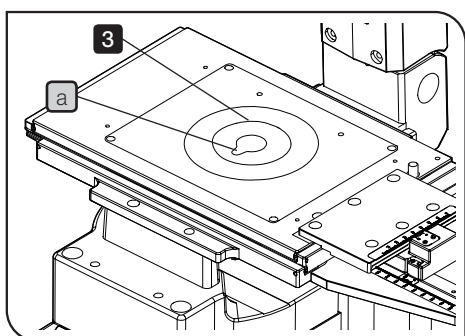
3 Fit the stage center plate into the stage.

CAUTION The stage center plate is designed very thin so that the objective will not hit it when the revolving nosepiece is rotated. Do not subject the stage center plate to impact or excessive force, as this may deform it.

⊙ To facilitate confirmation of the objective front lens position and application of oil for oil immersion objectives, rotate the stage center plate so that the keyhole **a** will be facing the front.

4 Screw the stage clips IX-SCL into the threaded holes provided on the stage top surface as needed.

⊙ The stage IX3-SVR can be mounted by reversing the front and rear to implement a stage with knobs on the left position. In this case, the movement range will be decreased and it will be difficult to check scales.



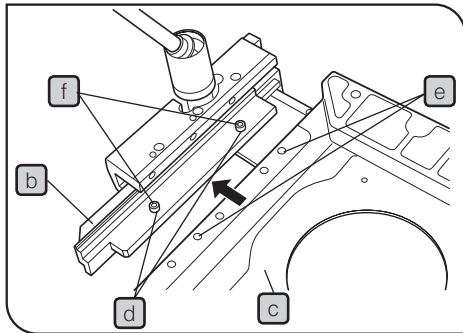
Attaching the sample holder

5 Remove the holder fixing screws (4 screws) by using the Allen wrench provided with IX3-SVR.

6 Remove the round stage center plate holder.

7 Set IX3-HOW in the center of the stage, and attach the holder fixing screws.

8 If you are using IX3-HOW or IX3-HO35D, set the sample holder in the center of IX3-HOW.



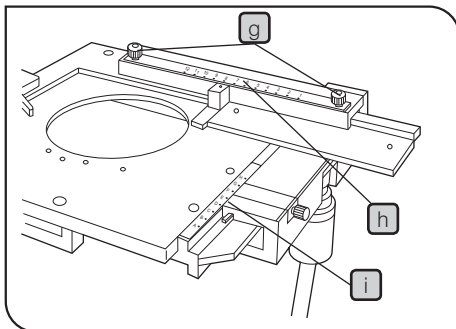
Mechanical Stage (IX-MVR)

Attaching Procedure

- 1 Invert the mechanical stage **b** and the plain stage IX2-SP **c** upside down.
- 2 Align the two mounting holes **d** on the mechanical stage with the mounting holes **e** on the plain stage. Insert the two clamping screws **f** and tighten them using the Allen screwdriver.

CAUTION

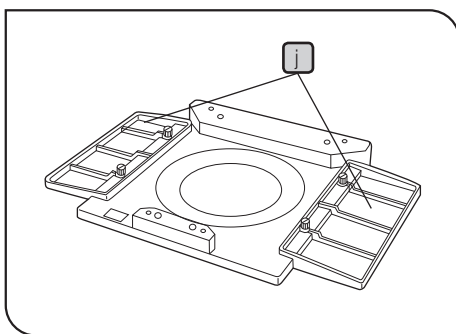
The mechanical stage may also be attached to the left side of the plain stage. However, this will interfere with the operation of the stage travel knobs, focusing knobs and the revolving nosepiece.



Attaching the scales

⊙ Scales for use with a 96-well microplate are provided with the mechanical stage.

- 1 Loosen the two clamping knobs **g** at the top of the stage's X-axis guide. Positioning the scale numerals correctly, place the X-axis scale **h** on the guide. Then tighten the clamping knobs.
- 2 For positioning the scale numerals correctly, place the Y-axis scale **i** on the top of the stage's Y-axis guide. The scale will be held in place magnetically.



Attaching the substages CK2-SS

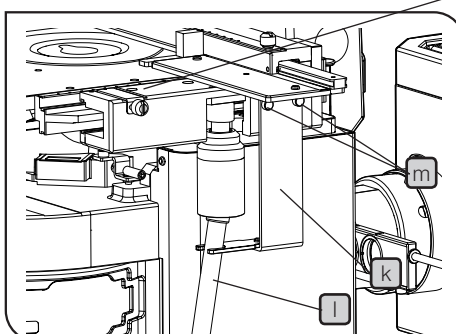
These plates can be attached in the same way as outlined in mechanical stage (IX-MVR) above.

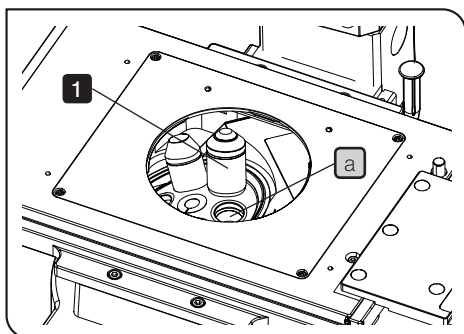
⊙ When the mechanical stage is not used, stage extender plates **j** can be attached at both sides of the plain stage.

Attaching the positioning bracket

⊙ A flexible stalk for the coaxial X-axis and Y-axis knobs is employed with the mechanical stage IX-MVR. These stages are provided with a positioning bracket that allows the tilt of the stalk to be adjusted to suit the observer's preference.

- 1 Insert the stalk **l** of the flexible X-axis/Y-axis knobs into the U-shaped notch on the positioning bracket **k**.
- 2 Using the Allen screwdriver, tighten the clamping screw **m** of the bracket lightly to the stage.
- 3 Align the stage's center alignment index line with the edge of the upper stage.
- 4 Slide the positioning bracket to and fro in order to determine the most convenient stalk angle.
- 5 Tighten the bracket's clamping screw firmly.



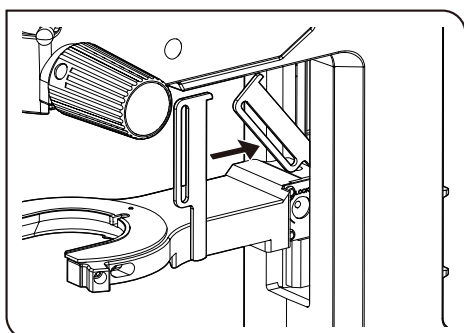


5 Attaching the Objectives

- 1 Remove the stage center plate and attach the objectives to the revolving nosepiece through the hole on the stage left by the plate.
- ⊙ Attach the objectives in such a manner that the magnification increases from low to higher powers in a clockwise direction.

CAUTION In the inverted microscope, the front lens of the objectives faces upward, and is more exposed to contamination than the objectives of upright microscopes. Therefore, if there are empty positions **a** in the nosepiece, attach the dust caps provided.

- 2 After all objective lenses are mounted, fit the stage center plate into the stage.

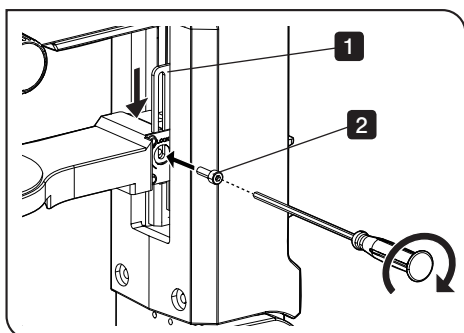


6 Attaching the Condenser Refocusing Stopper

only IX3-ILL

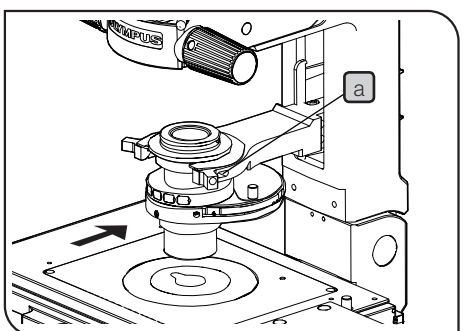
CAUTION If the condenser refocusing stopper is attached when the condenser attaching arm is placed at the lower position, the condenser may not be attached. Be sure to raise the condenser attaching arm before attaching the condenser refocusing stopper.

- ⊙ Mount the condenser refocusing stopper in the direction shown in the picture.



- 1 Insert the condenser refocusing stopper between the column and the condenser attaching arm.
- 2 Secure the condenser refocusing stopper with the fixing screw by using the Allen screwdriver provided with the microscope.

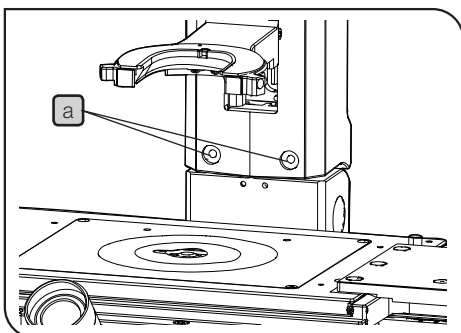
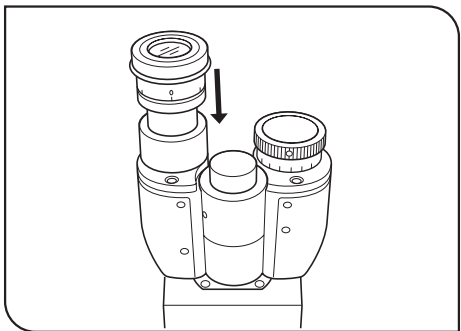
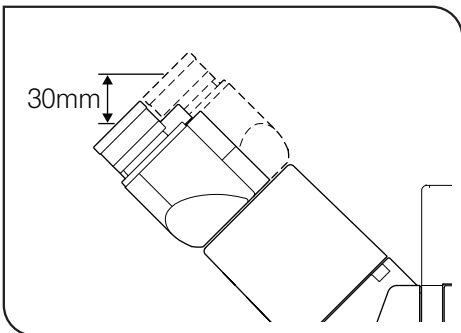
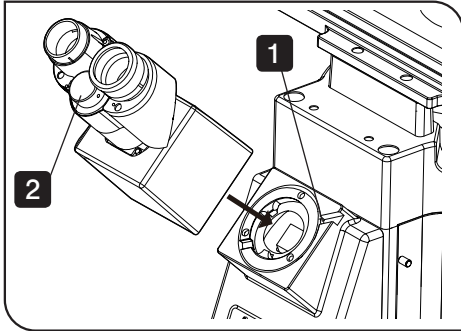
CAUTION Be careful not to drop the fixing screws.



7 Attaching the Condenser

only IX3-ILL

- 1 Loosen the condenser clamping screw **a**.
- 2 Fit the condenser into the mount dovetail on the condenser holder, and push in the condenser until its positioning pin fits into the positioning groove on the mount dovetail.
- 3 Tighten the condenser clamping screw **a**.



8 Attaching the Observation Tube

Binocular Tube (U-BI90, U-TBI90)

- 1 Using the Allen screwdriver, loosen the observation tube clamping screw on the observation tube mount to remove the cap.
- 2 Attach the circular dovetail mount of the observation tube into the observation tube mount, placing the observation tube so that the interpupillary distance scale numbers are seen right side up. Then tighten the clamping screw to clamp the observation tube.

⊙ Normally, the distance from the surface of the desk to the eye point is approximately 430 mm. If it is desirable to lower the eye point position, the observation tube (provided it is other than the U-TBI90) may be detached, turned 180° and then attached on the observation tube mount again. This lowers the eye point height by approximately 30 mm.

9 Attaching the Eyepieces

- 1 Remove the eyepieces' dust caps.
- 2 Insert the WHN10X-H eyepiece with helicoid into the eyepiece sleeve without helicoid (shown on the left in the figure).
- 3 Insert the WHN10X eyepiece without helicoid into the eyepiece sleeve with helicoid (shown on the right in the figure).

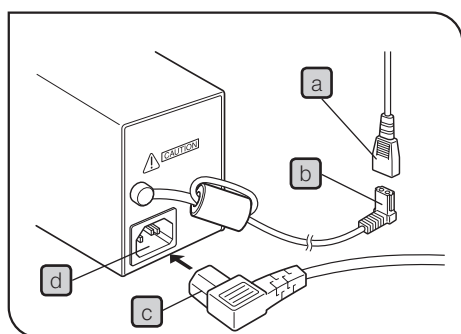
10 Using the Accessory Mounting Holes

- ⊙ Twelve threaded holes **a** are provided on the microscope for mounting accessories such as a micromanipulator.
- ⊙ Insert the cylindrical material (diameter: less than 11.5 mm; height: higher than 5 mm) between the microscope, and mount accessories.

CAUTION The applicable screws are M6 screws. The screws can be inserted into the microscope up to a depth of 10 mm. Select the screw length in accordance with this.

11 Connecting the Cables

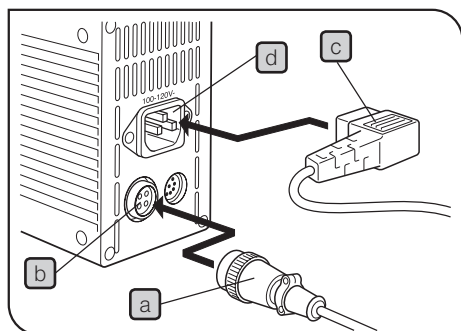
- CAUTION**
- Cables and cords are vulnerable when bent or twisted. Never subject them to excessive force.
 - Make sure that the main switch of the power supply is set to “OFF” before connecting cables.



TL4

TL4

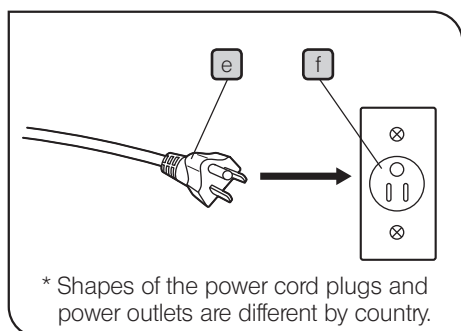
- 1 Connect the plug **a** of the illumination column to the connector **b** firmly.
- 2 Connect the power cord connector **c** to the connector **d** firmly.
- 3 Connect the power cord plug **e** to a wall outlet **f**.



TH4

TH4

- 1 Connect the plug **a** of the lamp housing to the connector **b** firmly.
- 2 Connect the power cord connector **c** to the connector **d** firmly.
- 3 Connect the power cord plug **e** to a wall outlet **f**.



TL4/TH4

10 LAMP HOUSING INSPECTION SHEET

IX2-ILL30 (30W Lamp socket)

- Study the instruction manual for the lamp housing before inspection.
- For safe use of the lamp housing, we recommend performing the following inspection periodically (every time you replace the lamp and at least every 6 months).
- The table below identifies the check items to be observed. Put (X) if not applicable or (✓) if applicable.
- If there is any (✓) mark noted, immediately stop use of the product, and contact Olympus for detailed inspections or replace the lamp housing.
- If you detect an abnormality other than that listed below or with other Olympus product, also stop the use of the product and contact Olympus for detailed inspections.
- Note that service, replacement and detailed inspections are charged after expiration of the warranty period.

If you have any questions, please contact Olympus.

Check items	Check results (Date)			
	/	/	/	/
1. More than 8 years have passed since original purchase or the total power ON time has exceeded 20,000 hours.				
2. Lamp does not light sometimes even though the main switch is set to on.				
3. Illumination flickers when you move the lamp cable or lamp socket.				
4. Scorching or burning odor is produced during use.				
5. Illumination still flickers after replacement with a new lamp bulb.				
6. Deformation, backlash, or looseness, etc. when you assemble the lamp socket.				
7. Extreme discoloration of the lamp socket connection terminal or lamp socket lamp bulb mount.				
8. Discoloration, deformation or cracking of the lamp socket.				
9. Melting, crack, deformation or solidification of the lamp cable or a wiring part.				
10. Increased frequency of servicing compared to similar devices put into use at the same time as the lamp socket.				

* When the Check Result columns become insufficient, copy this sheet.

IX3-ILL (100W Lamp housing)

- Study the instruction manual for the lamp housing before inspection.
- For safe use of the lamp housing, we recommend performing the following inspection periodically (every time you replace the lamp and at least every 6 months).
- The table below identifies the check items to be observed. Put (X) if not applicable or (√) if applicable.
- If there is any (√) mark noted, immediately stop use of the product, and contact Olympus for detailed inspections or replace the lamp housing.
- If you detect an abnormality other than that listed below or with other Olympus product, also stop the use of the product and contact Olympus for detailed inspections.
- Note that service, replacement and detailed inspections are charged after expiration of the warranty period.

If you have any questions, please contact Olympus.

Check items	Check results (Date)			
	/	/	/	/
1. More than 8 years have passed since original purchase or the total power ON time has exceeded 20,000 hours.				
2. Illumination flickers when you move the lamp cable or lamp housing.				
3. Lamp cable is unusually hot to the touch.				
4. Scorching or burning odor is produced during use.				
5. Deformation, backlash, or looseness, etc. when you assemble the lamp housing. (Impossibility of removing the top section of lamp housing when you attempt to replace the lamp, etc.)				
6. Discoloration, deformation or cracking of the lamp housing.				
7. Melting, crack, deformation or solidification of the lamp cable or a wiring part.				
8. Increased frequency of servicing compared to similar devices put into use at the same time as the lamp housing.				

* When the Check Result columns become insufficient, copy this sheet.

■ PROPER SELECTION OF THE POWER SUPPLY CORD

If no power supply cord is provided, please select the proper power supply cord for the equipment by referring to “ Specifications ” and “ Certified Cord ” below:

CAUTION: In case you use a non-approved power supply cord for Olympus products, Olympus can no longer warrant the electrical safety of the equipment.

Specifications

Voltage Rating	125V AC (for 100-120V AC area) or, 250V AC (for 220-240V AC area)
Current Rating	6A minimum
Temperature Rating	60°C minimum
Length	3.05 m maximum
Fittings Configuration	Grounding type attachment plug cap. Opposite terminates in molded-on IEC configuration appliance coupling.

Table 1 Certified Cord

A power supply cord should be certified by one of the agencies listed in Table 1 , or comprised of cordage marked with an agency marking per Table 1 or marked per Table 2. The fittings are to be marked with at least one of the agencies listed in Table 1. In case you are unable to buy locally in your country the power supply cord which is approved by one of the agencies mentioned in Table 1, please use replacements approved by any other equivalent and authorized agencies in your country.




















Country	Agency	Certification Mark	Country	Agency	Certification Mark
Argentina	IRAM		Italy	IMQ	
Australia	SAA		Japan	JET, JQA,	
Austria	ÖVE		Netherlands	KEMA	
Belgium	CEBEC		Norway	NEMKO	
Canada	CSA		Spain	AEE	
Denmark	DEMKO		Sweden	SEMKO	
Finland	FEI		Switzerland	SEV	
France	UTE		United Kingdom	ASTA BSI	
Germany	VDE		U.S.A.	UL	
Ireland	NSAI				

Table 2 HAR Flexible Cord

APPROVAL ORGANIZATIONS AND CORDAGE HARMONIZATION MARKING METHODS

Approval Organization	Printed or Embossed Harmonization Marking (May be located on jacket or insulation of internal wiring)		Alternative Marking Utilizing Black-Red-Yellow Thread (Length of color section in mm)		
			Black	Red	Yellow
Comite Electrotechnique Belge (CEBEC)	CEBEC	⟨HAR⟩	10	30	10
Verband Deutscher Elektrotechniker (VDE) e.V. Prüfstelle	⟨VDE⟩	⟨HAR⟩	30	10	10
Union Technique de l'Electricite' (UTE)	USE	⟨HAR⟩	30	10	30
Instituto Italiano del Marchio di Qualita' (IMQ)	IEMMEQU	⟨HAR⟩	10	30	50
British Approvals Service for Electric Cables (BASEC)	BASEC	⟨HAR⟩	10	10	30
N.V. KEMA	KEMA-KEUR	⟨HAR⟩	10	30	30
SEMKO AB Svenska Elektriska Materielkontrollanstalter	SEMKO	⟨HAR⟩	10	10	50
Österreichischer Verband für Elektrotechnik (ÖVE)	⟨ÖVE⟩	⟨HAR⟩	30	10	50
Danmarks Elektriske Materialkontroll (DEMKO)	⟨DEMKO⟩	⟨HAR⟩	30	10	30
National Standards Authority of Ireland (NSAI)	⟨NSAI⟩	⟨HAR⟩	30	30	50
Norges Elektriske Materielkontroll (NEMKO)	NEMKO	⟨HAR⟩	10	10	70
Asociacion Electrotecnica Y Electronica Espanola (AEE)	⟨UNED⟩	⟨HAR⟩	30	10	70
Hellenic Organization for Standardization (ELOT)	ELOT	⟨HAR⟩	30	30	70
Instituto Portages da Qualidade (IPQ)	np	⟨HAR⟩	10	10	90
Schweizerischer Elektro Technischer Verein (SEV)	SEV	⟨HAR⟩	10	30	90
Elektriska Inspektoratet	SETI	⟨HAR⟩	10	30	90

Underwriters Laboratories Inc. (UL)
Canadian Standards Association (CSA)

SV, SVT, SJ or SJT, 3 X 18AWG
SV, SVT, SJ or SJT, 3 X 18AWG

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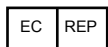


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